# **Sustainable Alternatives to Triton<sup>TM</sup> X-100 Detergent for Biomanufacturing: The Deviron® Detergent Portfolio**



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### Introduction

Virus Inactivation

## **Deviron® Detergents**

Viral safety is a major concern for biotherapeutic manufacturers.

- <u>Cell-based processes</u> may produce endogenous retroviral particles, and adventitious viruses can be introduced from contaminated source materials or during the manufacturing process.
- Human plasma-derived products are at risk of containing viruses, despite extensive screening of donation material.

**Detergent**-mediated **viral inactivation** is widely used in multiple biotherapeutic production processes as part of an overall virus safety strategy.

**Triton™ X-100** detergent is widely used.

- A degradation product of Triton<sup>™</sup> X-100 detergent is 4tert-octylphenol, an endocrine disruptor hazardous for the environment.
- Triton<sup>™</sup> X-100 detergent was classified as "**Substance of** Very High Concern SVHC" in the REACH (Registration, Evaluation, Authorization and Restriction of Chemicals) in 2017.
- The European Chemical Agency (ECHA) prohibited the **unauthorized use** of **Triton™ X-100** detergent in the **EU** in January **2021.**
- The biotherapeutic industry together is faced with the challenges of identifying, producing and implementing new alternatives.







Virus inactivation was assessed in mAb-containing CHO clarified harvest and human plasma matrices with XMuLV model.

The standard viral inactivation practice for **mAb processes** is described in the **ASTM E3042-16**. In this procedure the **detergent** concentration is **≥0.5 %, no solvent** (i.e., TnBP) and incubation time  $\geq 60$  min.

Plasma processes typically utilize 1.0 % detergent and **0.3 % solvent TnBP** and longer incubation time (4-6 h). Deviron<sup>®</sup> C16 and Deviron<sup>®</sup> 13-S9 detergents demonstrate effective viral inactivation (LRV > 5) in all conditions tested. Additional data with PRV and BVDV viruses are available.





Both Deviron<sup>®</sup> detergents were excellent in lysing HEK293 and Sf-RVN<sup>®</sup> cells (data not shown, microscopic images show complete cell lysis). Both Deviron<sup>®</sup> detergents were comparable or even better than the benchmarks.



Both Deviron<sup>®</sup> detergents were as efficient as the benchmarks in releasing capsids and the viral genome titers were comparably high.



The infectivity of the AAV is a crucial parameter for final products. Both of the Deviron<sup>®</sup> detergents preserve infectivity of the AAV after the lysis, as evaluated with the transduction unit assay.

The Deviron<sup>®</sup> detergents are an advantageous alternative to the benchmarks, combining efficacy, sustainability, and easier handling.

**Removal of Deviron<sup>®</sup> detergents in Downstream Process** Effective detergent removal from a process stream needs to be achieved by downstream steps. Removal was assessed with several different chromatographic resins.

#### Minimum criteria for alternative detergent candidates

![](_page_0_Figure_32.jpeg)

We offer a **portfolio of detergents**, the **Deviron**<sup>®</sup> detergents, to meet different application and process requirements.

### **Deviron® Detergents**

	Deviron <sup>®</sup> C-16 detergent	Deviron <sup>®</sup> 13-S9 detergent
Chemical name	N,N-Dimethyltetradecylamin- N-oxide	Alcohols, C11-15-secondary, ethoxylated
CAS number	3332-27-2	68131-40-8

#### **Cell Lysis**

HEK293 and Sf-RVN<sup>®</sup> cells were cultivated according to their type, suspension or adherent, in the appropriate cell culture media.

HEK293 cells were transfected after one day of culture with the selected plasmid polyethylenimine (PEI) complexation and Sf-RVN<sup>®</sup> cells infected with Baculovirus at the time of seeding. After cultivation, the cells were lysed.

#### Detergent: 0.5 % wt.

**Nuclease: 25 U/mL Benzonase®** endonuclease with 2 mM

#### **Deviron® C16 detergent**

Columns tested	Type of resin	n Removal	We recommend
Protein A	Eshmuno <sup>®</sup> A	99.6 %	inactivation with Deviron <sup>®</sup> C16 detergent prior to protein A
CEX (pH 6)	Eshmuno <sup>®</sup> S	43-64 %	affinity capture step
AEX (bind/elute mode pH 9)	Fractogel <sup>®</sup> TMAE Hicap	99.8 %	
AEX (flow through mode pH 9)	Fractogel <sup>®</sup> TMAE Hicap	Not suitable	
• pH< 8 • pH>8.	$3.9 \rightarrow \text{net position}$	ve charge	Resin selection depends on
	.9 → net negati	ve charge	<ul> <li>Unit operation pH</li> </ul>
Deviron	.9 → net negati ® <b>13-S9 dete</b>	ve charge ergent	<ul> <li>Unit operation pH</li> <li>Protein pI</li> </ul>
Deviron Columns tested	.9 → net negati <sup>®</sup> 13-S9 dete Type of resin	ve charge ergent Removal	<ul> <li>Unit operation pH</li> <li>Protein pI</li> </ul>
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Deviron Columns tested Protein A CEX (pH 6)	.9 → net negati <b>® 13-S9 dete</b> Type of resin Eshmuno® A Eshmuno® S	ve charge ergent Removal 100 % 99 %	<ul> <li>Unit operation pH</li> <li>Protein pI</li> </ul>

Surfactant	Zwitterionic (pI 8.9)	Non-ionic
СМС	0.002-0.003 wt % (24 °C)	0.005 wt % (24 °C)
Form	30 % wt. water solution	Pure substance (100 % wt.)
Biodegradability (OECD 301B)	Readily biodegradable	Readily biodegradable
Toxicology report	Available	Available
Quality marker	ISO9001	IPEC-PQG GMP
Documentation package	Emprove <sup>®</sup> Evolve MQ400	Emprove <sup>®</sup> Expert MQ500
Viral inactivation efficiency	Yes >5 LRV	Yes >5 LRV
Cell lysis efficiency	Yes	Yes
Endotoxin removal for plasmid purification	Yes	Yes
<b>Detection method</b>	HPLC-ELSD method available	HPLC-ELSD method available

Lysis time: 2 h

Temperature: 37 °C

Following incubation, total cell count and viability values determined. Virus-containing supernatant was clarified by centrifugation. The supernatant was analyzed to determine physical titer and genome titer. After detergent removal, the AAV infectivity was measured.

![](_page_0_Figure_49.jpeg)

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![](_page_0_Figure_52.jpeg)

MgCl<sub>2</sub>