

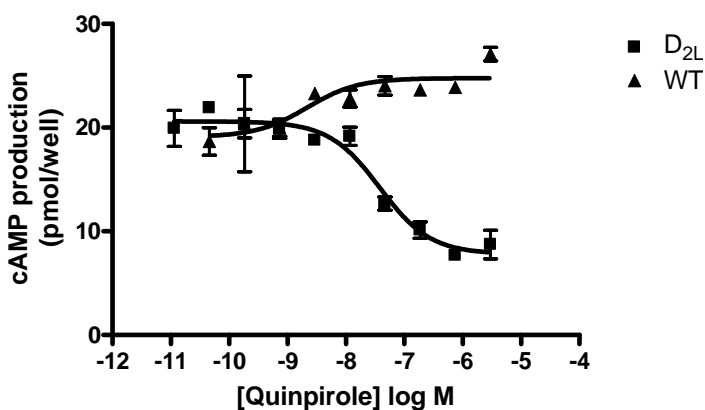


## ChemiScreen™ cAMP -OPTIMIZED STABLE CELL LINE HUMAN RECOMBINANT D<sub>2L</sub> DOPAMINE RECEPTOR

**CATALOG NUMBER:** HTS039C2      **QUANTITY:** 2 vials, 1 mL per vial  
**LOT NUMBER:**      **CONCENTRATION:** 2 x 10<sup>6</sup> cells/mL

**BACKGROUND:** Dopamine is a catecholamine neurotransmitter that functions in the CNS to control locomotor, cognitive, emotional and neuroendocrine processes, and in the periphery to modulate cardiovascular, renal and gastrointestinal processes. The biological activities of dopamine are mediated by a family of five GPCRs. The D<sub>1</sub> and D<sub>5</sub> subtypes couple to G<sub>s</sub> to increase intracellular cAMP, whereas the D<sub>2</sub>, D<sub>3</sub> and D<sub>4</sub> subtypes couple to G<sub>i</sub> to reduce cAMP (Missale *et al.*, 1998). The D<sub>2</sub> dopamine receptors have been of particular clinical interest due to their regulation of prolactin secretion and their affinity for antipsychotic drugs. The D<sub>2</sub> receptor exists as two alternatively spliced isoforms differing in the insertion of a stretch of 29 amino acids in the third intracellular loop (D<sub>2S</sub> and D<sub>2L</sub>) (Giros *et al.*, 1989; Grandy *et al.*, 1989). Millipore's cloned human D<sub>2L</sub>-expressing cell line is made in the CHO host, which supports optimal levels of recombinant D<sub>2L</sub> expression for robust agonist-induced cAMP signal. Thus, the cell line is an ideal tool for screening for agonists and antagonists at the D<sub>2L</sub> Receptor.

**APPLICATIONS:** Assay for inhibition of forskolin-induced cyclic AMP



**Figure 1.** Cyclic AMP assay with D<sub>2L</sub>-expressing CHO cell line. D<sub>2L</sub>-expressing CHO cells and wild-type CHO (WT) were preincubated in 1 mM IBMX for 5 min, then exposed to ligand in the presence of 10  $\mu$ M forskolin for another 15 min at 37°C. Cells were lysed and cAMP levels were determined with Millipore's cAMP HTS immunoassay kit (catalog # 17-418).



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**Table I.** Comparison of EC<sub>50</sub> values of D<sub>2L</sub>-expressing CHO cells with values described in the literature.

ligand	assay	potency (nM)	Reference
Quinpirol	cAMP	EC <sub>50</sub> = 38	Figure 1
Quinpirol	cAMP	EC <sub>50</sub> = 14	Moreland <i>et al.</i> (2004)

HOST CELLS: CHO-K1 cells

TRANSFECTION: Plasmid pcDNA3 containing DRD2 long isoform cDNA encoding D<sub>2L</sub> (Accession Number: NM\_000795; see CODING SEQUENCE below). The stable clonal cell line was selected by resistance to geneticin, followed by limited dilution cloning. The cell line was tested and found to have equivalent EC<sub>50</sub> and signal at 1, 3 and 6 weeks of continuous culture.

### PRESENTATION:

Cells are frozen at 2 x 10<sup>6</sup> cells/mL in 90% fetal bovine serum/10% DMSO. Cell line tests negative for mycoplasma.

### STORAGE/HANDLING:

1. Immediately upon receipt, thaw cells or place cells in liquid nitrogen.
2. Thaw cells rapidly by removing from liquid nitrogen and immediately immersing in a 37°C water bath. Immediately after ice has thawed, sterilize the exterior of the vial with 70% ethanol. Transfer contents of the vial to a T75 flask containing growth media. Place the flask in a humidified incubator at 37°C with 5% CO<sub>2</sub>.
3. After 8-24 h, all live cells will be attached. Viability of the cells is expected to be 50-80%. At this time, replace media to remove residual DMSO, and return to incubator.
4. When cells are approximately 80% confluent, passage the cells as follows: Remove media and wash once with HBSS without Ca<sup>++</sup> and Mg<sup>++</sup> (10 mL/T75). Add 0.05% trypsin/0.2 g/L EDTA (1 mL/T75) and place in humidified incubator at 37°C with 5% CO<sub>2</sub> until cells begin to round up and detach (5-10 minutes). Gently rap the side of the flask to dislodge the cells. Neutralize trypsin by addition of 4 mL CHO Growth Media per 1 mL trypsin.
5. Cells are typically passaged 1:10 every 3-4 days. Passaging ratio may be varied according to requirements of the investigator.
6. Frozen stocks of cells should be prepared at the earliest passage possible after thawing, as follows: Count detached cells (prepared as in Step 4). Centrifuge cells at 200 x g for 5 min. Resuspend cells at 5 x 10<sup>6</sup> cells/mL in CHO Freezing Media (cell densities of 2-10 x 10<sup>6</sup> are also acceptable if necessary). Dispense 1 mL aliquots into cryopreservation vials. Freeze the cells by a controlled rate process, such as in an isopropanol-jacketed container placed at -70°C overnight. Store the vials in liquid nitrogen.
7. Use of cells immediately after thawing is feasible for some cell lines and is being



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further validated. Some cell lines may need to be passaged at least once after thawing prior to use in calcium flux assays. Cells should be resuspended in CHO Plating Media for plating for calcium assay.

### MEDIA:

#### CHO Growth Media:

F12-K containing 2 mM L-glutamine (Invitrogen 21127)  
10% heat-inactivated FBS  
1x Pen-Strep (from 100x stock, Millipore TMS-AB2-C)  
250µg/mL Genetecin/G-418

#### CHO Plating Media:

F12-K containing 2 mM L-glutamine (Invitrogen 21127)  
10% heat-inactivated FBS  
1x Pen-Strep (from 100x stock, Millipore TMS-AB2-C)

#### CHO Freezing Media:

90% heat-inactivated FBS  
10% DMSO (cell culture grade)

### EXAMPLE CYCLIC AMP ASSAY CONDITIONS:

1. Cells propagated for screening should be maintained and seeded at less than 90% confluency. Trypsinize cells as above and seed cells in 96-well tissue culture plate at 50,000 cells/well in CHO Plating Media. Incubate plate overnight in a humidified incubator at 37°C with 5% CO<sub>2</sub>.
2. Remove media from the cells and add 50ul/well of cAMP assay buffer (HBSS containing calcium and magnesium, with 10 mM HEPES) containing 2mM IBMX. Incubate cells in a humidified 37°C/5% CO<sub>2</sub> incubator for 5 min.
3. Add 50ul/well of cAMP assay buffer by itself or containing 2x final concentration of desired concentration of control or testing compounds and 20 µM forskolin. Incubate cells in a humidified 37°C/5% CO<sub>2</sub> incubator for 15 min.
4. Terminate the reaction by adding 100ul/well of lysis buffer from cAMP HTS immunoassay kit (Millipore 17-418), and perform cAMP quantitation according to the kit instructions.

### REFERENCES:

Grandy DK *et al.* (1989) Cloning of the cDNA and gene for a human D<sub>2</sub> dopamine receptor. *Proc. Natl. Acad. Sci. USA* 86:9762-6.

Giros B *et al.* (1989) Alternative splicing directs the expression of two D<sub>2</sub> dopamine receptor isoforms. *Nature* 342:923-6.

Missale C *et al.* (1998) Dopamine receptors: from structure to function. *Physiol. Rev.* 78: 189-225.

Moreland RB *et al.* (2004) Comparative pharmacology of human dopamine D<sub>2</sub>-like receptor stable cell lines coupled to calcium flux through G<sub>αq05</sub>. *Biochem. Pharmacol.* 68:



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761-772.

## CODING SEQUENCE:

```
1 - ATG GAT CCA CTG AAT CTG TCC TGG TAT GAT GAT GAT CTG GAG AGG CAG AAC TGG AGC CGG - 60
1 - M D P L N L S W Y D D D L E R Q N W S R - 20

61 - CCC TTC AAC GGG TCA GAC GGG AAG GCG GAC AGA CCC CAC TAC AAC TAC TAT GCC ACA CTG - 120
21 - P F N G S D G K A D R P H Y N Y A T L - 40

121 - CTC ACC CTG CTC ATC GCT GTC ATC GTC TTC GGC AAC GTG CTG GTG TGC ATG GCT GTG TCC - 180
41 - L T L L I A V I V F G N V L V C M A V S - 60

181 - CGC GAG AAG GCG CTG CAG ACC ACC ACC AAC TAC CTG ATC GTC AGC CTC GCA GTG GCC GAC - 240
61 - R E K A L Q T T T N Y L I V S L A V A D - 80

241 - CTC CTC GTC GCC ACA CTG GTC ATG CCC TGG GTT GTC TAC CTG GAG GTG GTA GGT GAG TGG - 300
81 - L L V A T L V M P W V V Y L E V V G E W - 100

301 - AAA TTC AGC AGG ATT CAC TGT GAC ATC TTC GTC ACT CTG GAC GTC ATG ATG TGC ACG GCG - 360
101 - K F S R I H C D I F V T L D V M M C T A - 120

361 - AGC ATC CTG AAC TTG TGT GCC ATC AGC ATC GAC AGG TAC ACA GCT GTG GCC ATG CCC ATG - 420
121 - S I L N L C A I D R Y T A V A M P M - 140

421 - CTG TAC AAT ACG CGC TAC AGC TCC AAG CGC CGG GTC ACC GTC ATG ATC TCC ATC GTC TGG - 480
141 - L Y N T R Y S S K R R V T V M I S I V W - 160

481 - GTC CTG TCC TTC ACC ATC TCC TGC CCA CTC CTC TTC GGA CTC AAT AAC GCA GAC CAG AAC - 540
161 - V L S F T I S C P L L F G L N A D Q N - 180

541 - GAG TGC ATC ATT GCC AAC CCG GCC TTC GTG GTC TAC TCC TCC ATC GTC TCC TTC TAC GTG - 600
181 - E C I I A N P A F V V Y S S I V S F Y V - 200

601 - CCC TTC ATT GTC ACC CTG CTG GTC TAC ATC AAG ATC TAC ATT GTC CTC CGC AGA CGC CGC - 660
201 - P F I V T L L Y I K I Y I V L R M R - 220

661 - AAG CGA GTC AAC ACC AAA CGC AGC AGC CGA GCT TTC AGG GCC CAC CTG AGG GCT CCA CTA - 720
221 - K R V N T K R S S R A F R A H L R A P L - 240

721 - AAG GGC AAC TGT ACT CAC CCC GAG GAC ATG AAA CTC TGC ACC GTT ATC ATG AAG TCT AAT - 780
241 - K G N C T H P E D M K L C T V I M K S N - 260

781 - GGG AGT TTC CCA GTG AAC AGG CCG AGA GTG GAG GCT GCC CGG CGA GCC CAG GAG CTG GAG - 840
261 - G S F P V N R R R V E A A R R A Q E L E - 280

841 - ATG GAG ATG CTC TCC AGC ACC AGC CCA CCC GAG AGG ACC CGG TAC AGC CCC ATC CCA CCC - 900
281 - M E M L S S T S P P E R T R Y S P I P P - 300

901 - AGC CAC CAC CAG CTG ACT CTC CCC GAC CCG TCC CAC CAT GGT CTC CAC AGC ACT CCT GAC - 960
301 - S H H Q L T L P D P S H H G L H S T P D - 320

961 - AGC CCC GCC AAA CCA GAG AAG AAT GGG CAT GCC AAA GAC CAC CCC AAG ATT GCC AAG ATC - 1020
321 - S P A K P E K N G H A K D H P K I A K I - 340

1021 - TTT GAG ATC CAG ACC ATG CCC AAT GGC AAA ACC CGG ACC TCC CTC AAG ACC ATG AGC CGT - 1080
341 - F E I Q T M P N G K T R T S L K T M S R - 360

1081 - AGG AAG CTC TCC CAG CAG AAG GAG AAG AAA GCC ACT CAG ATG CTC GCC ATT GTT CTC GGC - 1140
361 - R K L S Q Q K E K K A T Q M L A I V L G - 380

1141 - GTG TTC ATC ATC TGC TGG CTG CCC TTC TTC ATC ACA CAC ATC CTG AAC ATA CAC TGT GAC - 1200
381 - V F I I C W L P F F I T H I L N I H C D - 400

1201 - TGC AAC ATC CCG CCT GTC CTG TAC AGC GCC TTC ACG TGG CTG GGC TAT GTC AAC AGC GCC - 1260
401 - C N I P P V L Y S A F T W L G Y V N S A - 420

1261 - GTG AAC CCC ATC ATC TAC ACC ACC TTC AAC ATT GAG TTC CGC AAG GCC TTC CTG AAG ATC - 1320
421 - V N P I I Y T T F N I E F R K A F L K I - 440

1321 - CTT CAC TGC TGA
441 - L H C *
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