

# 3dGRO™ Human Colon Organoid Expansion Medium

Stem Cell Media

Cat. # **SCM304**

FOR RESEARCH USE ONLY.  
NOT FOR USE IN DIAGNOSTIC PROCEDURES.  
NOT FOR HUMAN OR ANIMAL CONSUMPTION.

pack size: 50 ml

Store at -20°C



Data Sheet

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## Description

3dGRO™ Human Colon Organoid Expansion Medium is a complete ready-to-use serum-free medium for the expansion and long-term culture of human colon organoids.

The medium has been validated for use with 3dGRO™ Human iPSC Derived Colon Organoids (Cat. No. SCC300) and is expected to also work with patient-derived colon organoids. Organoids propagated in the medium express colon-specific markers including the posterior hindgut marker CDX2,  $\alpha$ -carbonic anhydrase II (CA-II),  $\alpha$ -carbonic anhydrase IV (CA-IV), and goblet cell markers Mucin-2 and Mucin-5B.

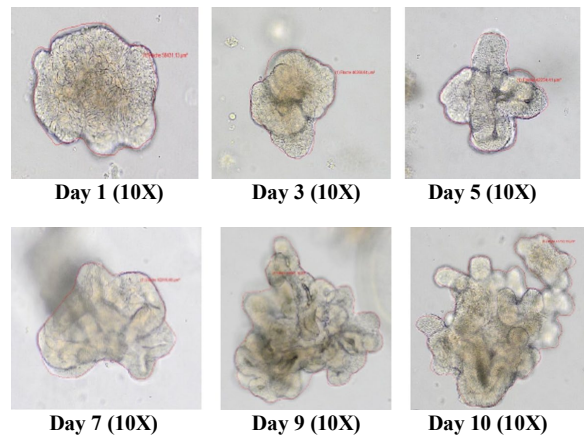
## Storage and Handling

Upon receipt, store at -20°C. When ready to use, thaw overnight at 2-8°C. Once thawed, mix thoroughly. Use immediately and store at 2-8°C for up to 1 week. Do not re-freeze. Unused aliquots may be stored at -20°C until the expiry date.

## Quality Control Testing

- Appearance (Color): Clear/No Particulates (Red Liquid)
- Osmolality: 350-375 mOsm
- pH: 7.0 – 7.4
- Sterility Tested: No Growth/Pass
- Endotoxin: <2 EU/mL
- Mycoplasma: Negative
- Functional Assay: Thaw and culture of human colon organoids for 2 passages.

## Representative Images



**Figure 1. Morphology and growth of human colon organoids.** Human colonic organoids were expanded in 3dGRO™ Human Colon Organoid Expansion Medium (SCM304) over a 10-day period and an increase in overall mean length and area was observed.

## References

1. Clevers H et al. (2011) Long-term expansion of epithelial organoids from human colon, adenoma, adenocarcinoma, and Barrett's epithelium. *Gastroenterology* 141(5): 1762-1772.
2. Spence JR et al. (2011) Directed differentiation of human pluripotent stem cells into intestinal tissue in vitro. *Nature* 470 (7332): 105-109.
3. Múnera JO et al. (2017) Differentiation of human pluripotent stem cells into colonic organoids via transient activation of BMP signaling. *Cell Stem Cell* 21(1): 51-64.
4. Crespo M et al. (2017) Colonic organoids derived from human induced pluripotent stem cells for modeling colorectal cancer and drug testing. *Nat Med.* 23(7): 878-884.

**SPECIES LEGEND:** H Human Ca Canine M Mouse R Rat Rb Rabbit B Bovine P Porcine WR Most Common Vertebrates

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**Important Notes before Starting:**

- The term “domes” refer to organoids that are 3D encapsulated in Growth Factor Reduced (GFR) Matrigel (Corning Cat. No. 356231).
- While not necessary, it is highly recommended that organoids are cultured in medium containing penicillin and streptomycin to prevent contamination that may be introduced during the long culture process.

**Reagents Required but Not Provided**

Products	Catalog #	Notes
Matrigel® Growth Factor reduced (GFR) Basement Membrane Matrix	Corning 356231	Thaw and maintain on ice. Make 1 mL aliquots. Aliquot the amount needed and maintain on ice. Store unused aliquots at -20°C.
3dGRO™ Human iPSC Derived Colon Organoids or patient derived colon organoids	SCC300	
ROCK Inhibitor, Y-27632	SCM075	Make a 10 mM or 1000X stock by reconstituting 5 mg with 1.47 mL sterile water. Aliquot and store long-term at -20°C.
Penicillin-Streptomycin Solution (100X)	TMS-AB2-C	Aliquot & store long-term at -20°C
24-well tissue culture treated plates	ThermoFisher 142475	
3dGRO™ Organoid Dissociation Reagent	SCM300	
3dGRO™ Organoid Freeze Medium	SCM301	

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Cat # SCM304

### Thawing Organoids into 24-well plates:

1. Before thawing, prepare sufficient Growth Factor Reduced (GFR) Matrigel for 12 domes at 25  $\mu$ L per dome + 5% overage (315  $\mu$ L total).  
**Note:** GFR Matrigel will gel at room temperature; maintain on ice at all times.
2. Prepare 3dGRO™ Expansion Medium (Cat. No. SCM304) supplemented with 1X Penicillin/Strep and 10  $\mu$ M ROCKi. For example, add 250  $\mu$ L of 100X Pen/Strep solution and 25  $\mu$ L of 10 mM ROCKi solution (10  $\mu$ M final) to 25 mL 3dGRO™ Expansion Medium. For media containing ROCKi, prepare fresh on the day of media change.
3. Remove the vial of cryopreserved organoids from liquid nitrogen storage and quickly thaw in a 37°C water bath. Closely monitor until only small ice crystals remain. Quickly remove the vial from the water bath. **IMPORTANT: Do not vortex the vial or leave in the water bath for too long.** Disinfect the outside of the vial with 70% ethanol or isopropanol.
4. In a laminar flow hood, use a 1 or 2 mL pipette to transfer the organoid suspension to a sterile 15 mL conical tube containing 9 mL 3dGRO™ Expansion Medium supplemented with 1X Pen/Strep and 10  $\mu$ M ROCKi (from step 2) Be careful not to introduce any air bubbles during the transfer process.
5. Centrifuge at 1100 rpm for 5 minutes at 4°C. Carefully aspirate the supernatant by connecting a P-200 pipette tip to the end of an aspirating pipette. Be careful not to aspirate the organoid pellet.
6. Immediately transfer 315  $\mu$ L of the ice-cold GFR Matrigel to the organoid pellet. GFR Matrigel may be viscous; ensure even mixing of the organoid suspension with GFR Matrigel by pipetting up and down several times with a P-1000 pipette. Be careful not to introduce air bubbles during pipetting. Place on ice for 3-5 minutes to cool down the organoid suspension.  
**TIP:** Set the pipette to 290  $\mu$ L instead of 315  $\mu$ L and resuspend the organoid pellet as quickly as you can. This will minimize air bubbles during pipetting.
7. Set a P-200 pipette to 25  $\mu$ L. Swirl the 15 mL conical tube containing the organoid Matrigel suspension to mix. Dispense 25  $\mu$ L of the organoid suspension into the center of each well of a 24-well plate. See Figure 3A. **NOTE: Do this as quickly as possible to prevent gelling of the organoid suspension.** Total number of domes = 12. Minimize air bubbles during pipetting.
8. Incubate in a 37°C, 5% CO<sub>2</sub> humidified incubator for 10 minutes. This will allow sufficient time for the organoid suspension to form a solid 3D "dome". See Figure 3A and 2B.
9. Gently add 1 mL of 3dGRO™ Expansion Medium supplemented with 1X Pen/Strep and 10  $\mu$ M ROCKi into each well containing the organoid domes. To avoid disturbing the domes, dispense the media onto the side of the wells.
10. Incubate in a 37°C, 5% CO<sub>2</sub> humidified incubator overnight.
11. Next day, inspect the organoids with a bright-field microscope. Live organoids should be rounded in shape and not fragmented (dead). Replace with freshly made 3dGRO™ Expansion Medium supplemented with 1X Pen/Strep and 10  $\mu$ M ROCKi. Incubate at 37°C overnight.
12. On the 2<sup>nd</sup> day after thaw, replace with fresh 3dGRO™ Expansion Medium supplemented with 1X Pen/Strep and 10  $\mu$ M ROCKi. Incubate at 37°C overnight. **Note:** ROCKi is only added for the first 2 days after thaw to enhance cell viability. Thereafter, ROCKi is no longer necessary.
13. Replace the media every other day with 3dGRO™ Expansion Medium supplemented with 1X Pen/Strep. **Note.** Media should NOT contain ROCKi.
14. Organoids may be passaged every 10 -12 days of culture using the 3dGRO™ Organoid Dissociation Reagent (Cat No. SCM300). Do not exceed 12 days before passaging.

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