



- Tissue culture flasks
- Hemacytometer

Procedure:

To establish cultures from cryopreserved cells, first ensure that adequate equipment and reagents are available to perform the necessary procedures in a timely manner. Prepare a sufficient volume of growth medium using Pancreatic Cell Culture Supplement (SCR015) & Pancreatic Cell Culture Medium (SCR016) according to our instructions (refer to SCR015 or SCR016 product datasheets).

1. Remove the desired number of vials containing cells from storage and rapidly transfer to a 37°C water bath. Provide continuous agitation, e.g., swirling, to the vial while it is submerged in the 37°C water bath. Use proper safety precautions for handling extremely cold materials including gloves and eye protection. Continue with agitation until the cells are completely thawed and no ice remains within the cell suspension, about 1 to 2 minutes. *Maximum cell viability is dependent on rapid and complete thawing of frozen cells.*
2. Transfer the thawed cells to a 15 mL centrifuge tube, add approximately 10 mLs of PBS to the centrifuge tube, agitate and then centrifuge at 460 x g for 10 minutes. Decant the supernatant as completely as possible and resuspend the cells in a total volume of 1 mL of growth medium by repetitive elutriation using a 1 mL pipette. This procedure washes out the cryopreservative used to freeze the cells. Count cells by a suitable method including a hemacytometer or automated cell counting device
3. Cultures established from frozen cell stocks require adaptation to achieve maximum growth rates and this usually requires about two to three successive passages. We suggest establishing the initial culture of LT2 cells derived from frozen stock at a plating density of about 15,000 cells/cm², followed by two successive passages where the cells are plated at 3,000 cells/cm². Rapid achievement of a homogeneous cell suspension within the tissue culture flask containing growth medium is necessary to establish uniform cell distribution within the culture. Following the initial plating, allow cells to incubate in 5% CO₂ at 37°C and monitor cell growth by visual inspection. When these cultures are 80% to 90% confluent, split and subculture the cells as described in the next section.

Subculture Procedures:**Reagents & Equipment required:**

- Accutase™ dissociation reagent (SCR005)
- 1 x PBS equilibrated to room temperature or 37°C
- Pancreatic Cell Culture Supplement (SCR015)
- Pancreatic Cell Culture Medium (SCR016)
- Growth medium: SCR016 supplemented with SCR015
- Tissue Culture Flasks (T-25 or T-75)
- Hemacytometer

Procedure:

1. Equilibrate sufficient PBS and Accutase™ to 37°C. Wash each flask three times with PBS (5 mLs per T-25 per wash or 15 mLs per T-75 per wash) and then add Accutase™



- (2 mLs/T-25 or 6 mLs/T-75) and incubate within the cell culture incubator for 10 to 15 minutes. Visualize the culture and assure complete detachment of cells by rapping the flask firmly on a solid surface.
2. Transfer the dissociated cells to a centrifuge tube and combine this with a PBS wash of the flask (5mL/T-25; 10 mL/T75). Centrifuge for 10 mins at 460 x g and remove the supernatant.
 3. Resuspend the pelleted cells in 1 mL growth medium by repetitive elutriation using a 1 mL pipette. Maintain the cell suspension at 4°C prior to completion of the subculture process. Count the cells using an automated cell counter or hemacytometer. For automated counters, count in the size range 4.5 to 17.5 µm.
 4. Plate LT2 cells at 3,000 cells/cm² for routine passage by adding the appropriate volume of cell suspension to growth medium.

For optimal viability, complete the subculture process within 2-3 hours of dissociation by Accutase™ treatment. Fully adapted LT2 cells require 4 to 5 days to reach approximately 90% confluence. We recommend feeding every two to three days. For longer or shorter periods between subculture, cultures may be inoculated at lower or higher densities. Subculture at less than 1,500 cells/cm² should be avoided to maintain maximal growth rates. Our suggested procedures are provided as guidelines and may require adjustments within different laboratory environments. These cells undergo crisis if feeding is too infrequent or subculture is delayed beyond confluence. Cultures will recover from mild crisis within a few passages but severe crisis conditions may necessitate re-establishing cultures from frozen stock.

Cryopreservation Procedure:

Reagents & Equipment required:

- Pancreatic Cell Cryopreservation Medium (SCR017)
- RPMI-1640 containing 25 mM HEPES, pH 7.2 (SLM-140-B)
- Cryogenic vials

Procedure:

1. Add an equal volume of 2X Pancreatic Cell Cryopreservation Medium to a known volume of cell suspension in RPMI 1640 containing 25 mM HEPES, pH 7.2. It is preferable to freeze LT2 cells at a concentration of 2 to 5 million cells per mL.
2. Dispense into cryogenic vials using suitable volumes for the desired applications.
3. Freeze the cell suspension at a slow rate, approximately 1°C/minute by standard methods.
4. After complete freezing, transfer vials of cells to liquid N₂ containing Dewar flask preferably in the vapor phase for long-term storage at maximum viability

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GMO

This product contains genetically modified organisms.
Este producto contiene organismos genéticamente modificados.
Questo prodotto contiene degli organismi geneticamente modificati.
Dieses Produkt enthält genetisch modifizierte Organismen.
Ce produit contient organismes génétiquement des modifiés.
Dit product bevat genetisch gewijzigde organismen.
Tämä tuote sisältää geneettisesti muutettuja organismeja.
Denna produkt innehåller genetiskt ändrade organismer.



Important Note: *During shipment, small volumes of product will occasionally become entrapped in the seal of the product vial. For products with volumes of 200 μ L or less, we recommend gently tapping the vial on a hard surface or briefly centrifuging the vial in a tabletop centrifuge to dislodge any liquid in the container's cap.*

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PROCEDURES. NOT FOR HUMAN OR ANIMAL CONSUMPTION

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