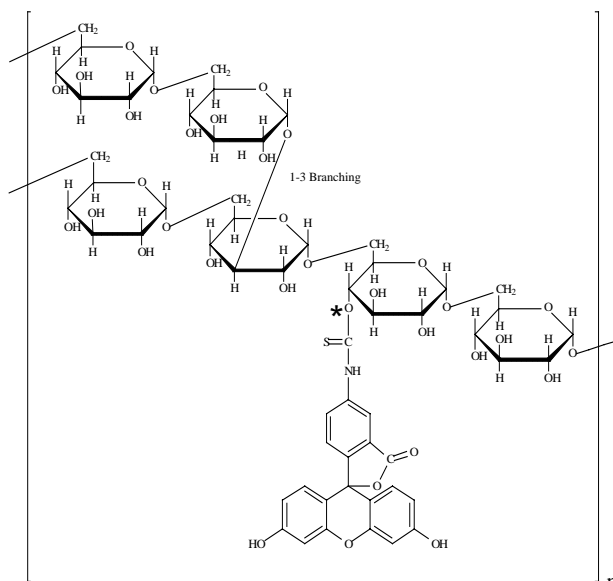


## Product Information

**FLUORESCCEIN ISOTHIOCYANATE-DEXTRAN**  
**Sigma Stock Nos. FD-4, FD-10S, FD-20S, FD-20,**  
**FD-40S, FD-40, FD-70S, FD-70, FD-150S, FD-150,**  
**FD-250S, FD-500S, and FD-2000S**



\* The site of attachment of FITC is assumed to be randomly associated with any free hydroxyl group

### PHYSICAL PROPERTIES:

Structure: Dextran is a polymer of anhydroglucose. It is composed of approximately 95% alpha-D-(166) linkages. The remaining (163) linkages account for the branching of dextran.<sup>1,2,3</sup> Conflicting data on the branch lengths implies that the average branch length is less than three glucose units.<sup>4,5</sup> However, other methods indicate branches of greater than 50 glucose units exist.<sup>6,7</sup> Native dextran has been found to have a molecular weight (MW) in the range of 9 million to 500 million.<sup>8,9,10</sup> Lower MW dextrans will exhibit slightly less branching<sup>4</sup> and have a more narrow range of MW distribution.<sup>11</sup> Dextrans with MW greater than 10,000 behave as if they are highly branched. As the MW increases, dextran molecules attain greater symmetry.<sup>7,12,13</sup> Dextrans with MW of 2,000 to 10,000, exhibit the properties of an expandable coil.<sup>12</sup> At MW below 2,000, dextran is more rod-like.<sup>14</sup> The MW of dextran is measured by one or more of the following methods: low angle laser light scattering,<sup>15</sup> size exclusion chromatography,<sup>16</sup> copper-complexation<sup>17</sup> and anthrone reagent<sup>18</sup> colorimetric reducing-end sugar determination and viscosity.<sup>12</sup>

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**PHYSICAL PROPERTIES:** (continued)

The approximate Stokes' radii for FITC-dextran are as follows;<sup>11</sup>

MW 4,000	Approx. 14 Angstroms
MW 10,000	Approx. 23 Angstroms
MW 20,000	Approx. 33 Angstroms
MW 40,000	Approx. 45 Angstroms
MW 70,000	Approx. 60 Angstroms
MW 150,000	Approx. 85 Angstroms

FITC is conjugated randomly to hydroxyl groups of dextran at a frequency of 0.003 to 0.02 moles of FITC per mole of glucose.

**FLUORESCENT PROPERTIES:**

The excitation maximum of FITC-dextran is 490 nm. The emission maximum is 520 nm.<sup>11</sup> The fluorescence increases with pH and is optimal at pH 8 and above.<sup>11</sup>

**SYNONYM:** FITC-Dextran

**METHOD OF PREPARATION:**

Sigma dextrans are derived from *Leuconostoc mesenteroides*, strain B 512. Various MW are produced by limited hydrolysis and fractionation. Our supplier's exact methods are held proprietary. Fractionation can be accomplished by size exclusion chromatography<sup>16</sup> or ethanol fractionation, where the largest MW dextrans precipitate first.<sup>19</sup> The FITC conjugation with hydroxyl groups of dextran is carried out in DMSO.<sup>20</sup>

**STABILITY/STORAGE AS SUPPLIED:**

Stored properly at 2-8°C and protected from light FITC-dextran powders should be stable for a minimum of two to three years.

**SOLUBILITY / SOLUTION STABILITY:**

Sigma typically tests the solubility of FITC dextrans in water at concentrations at or above 25 mg/ml. Solutions should be protected from light. In vivo, FITC-dextran is stable for more than 24 hours.<sup>21</sup>

**APPLICATIONS:**

FITC-dextran is used extensively in microcirculation and cell permeability research utilizing microfluorimetry.<sup>22,23</sup> FITC-dextran has been used to study plant cell wall porosity<sup>24</sup> and capillary permeability.<sup>25,26</sup> Plasma proteins have been shown not to bind to FITC-dextran.<sup>26</sup>

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**REFERENCES:**

1. Rankin, J.C. and Jeanes, A., *J. Am. Chem. Soc.*, 76, 4435 (1954).
2. Dimler, R.J. et al., *J. Am. Chem. Soc.*, 77, 6568 (1955).
3. Van Cleve, J.W. et al., *J. Am. Chem. Soc.*, 78, 4435 (1951).
4. Lindberg, B. and Svensson, S., *Acta. Chem. Scand.*, 22, 1907 (1968).
5. Larm, O. et al., *Carbohydr. Res.*, 20, 39 (1971).
6. Bovey, F.A., *J. Polym. Sci.*, 35, 167 (1959).
7. Senti, R.F. et al., *J. Polym. Sci.*, 17, 527 (1955).
8. Arond, L.H. and Frank, H.P., *J. Phys. Chem.*, 58, 953 (1954).
9. Elias, H.G., *Makromol. Chem.*, 33, 166 (1959).
10. Antonini, E. et al., *Biopolymers*, 2, 27 (1964).
11. Supplier's data.
12. Granath, K.A., *J. Colloid Sci.*, 13, 308 (1958).
13. Wales, M. et al., *J. Polym. Sci.*, 66, 101 (1979).
14. Gekko, K., *Am. Chem. Soc. Symposium Series*, 150, 415 (1981).
15. Allen, P. W., *Techniques of Polymer Characterization*, Butterworths Scientific Publications, p. 131 (1959).
16. Granath, K.A. and Flodin, P., *Makromol. Chem.*, 48, 160 (1961).
17. *Journal of Research of the National Bureau of Standards*, 50, 81 (1953).
18. Jermyn, M.A., *Anal. Biochem.*, 68, 332 (1975).
19. Ingelman, B. and Halling, M.S., *Ark. Kemi.*, 1, 61 (1949).
20. De Belder, A.N. and Gratath, K.A., *Carbohydrate Res.*, 30, 375 (1973).
21. Arfors, K.E. and Hint, H., *Microvasc. Res.*, 3, 440 (1971).
22. Arfors, K.E. and Rutili, G., *Microvasc. Res.*, 4, 466 (1972).
23. Arfors, K.E. and Rutili, G., Abstracts. VII Conference on Microcirculation, Aberdeen, p. 3 (1972).
24. Baron-Epal, O. et al., *Planta*, 175, 389 (1988).
25. Rutili, G. and Arfors, K. E., Abstracts. *Nordic Microcirculation Group Meeting*, Ustaoset (1973).
26. Rutili, G. and Arfors, K.E., Abstracts. VII Conference on Microcirculation, Aberdeen, p. 101 (1972).