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Product Information

PKACγ, active, GST-tagged, human PRECISIO® Kinase recombinant, expressed in *Sf*9 cells

Catalog Number **K3643** Storage Temperature –70 °C

Synonyms: KAPG, PKAr, cAPKr

Product Description

PKA C-gamma (PKAcγ) is a third isoform of the catalytic subunit of cAMP-dependent protein kinase. It was isolated from a human testis cDNA library and was clearly derived from a gene distinct from C-alpha and C-beta, and showed tissue-specific expression. Whereas, at the amino acid level C-alpha and C-beta show 93% homology, C-gamma shows only about 80% homology to both C-alpha and C-beta. The *PRKACG* gene is intronless, contains remnants of a poly(A) tail, is flanked by direct repeats, and is co-linear with the *PRKACA* gene.

This recombinant product was expressed by baculovirus in *Sf*9 insect cells using an N-terminal GST-tag. The gene accession number is NM 002732. It is supplied in 50 mM Tris-HCl, pH 7.5, with 150 mM NaCl, 0.25 mM DTT, 0.1 mM EGTA, 0.1 mM EDTA, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~65 kDa

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage/Stability

The product ships on dry ice and storage at -70 °C is recommended. After opening, aliquot into smaller quantities and store at -70 °C. Avoid repeated handling and multiple freeze/thaw cycles.

Figure 1.

SDS-PAGE Gel of Typical Lot:

≥70% (SDS-PAGE, densitometry)

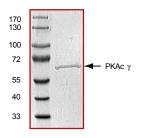
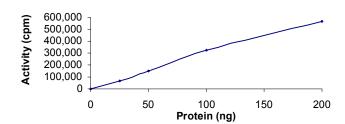


Figure 2.
Specific Activity of Typical Lot: 128–174 nmole/min/mg



Procedure

Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM MgCl₂, 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 ng/μl BSA solution.

Kinase Solution – Dilute the active PKAC γ (0.1 μ g/ μ l) with Kinase Dilution Buffer to the desired concentration. Note: The specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active PKAC γ kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 μ l aliquots at –20 °C.

 γ -³²P-ATP Assay Cocktail (250 μM) – Combine 5.75 ml of Kinase Assay Buffer, 150 μl of 10 mM ATP Stock Solution, 100 μl of γ -³²P-ATP (1 mCi/100 μl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Dissolve the synthetic peptide substrate (KRREILSRRPSYR) in water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

Kinase Assay

This assay involves the use of the ³²P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

- 1. Thaw the active PKAC γ , Kinase Assay Buffer, Substrate Solution, and Kinase Dilution Buffer on ice. The γ - 32 P-ATP Assay Cocktail may be thawed at room temperature.
- 2. In a pre-cooled microcentrifuge tube, add the following solutions to a volume of 20 μl:

10 ul of Kinase Solution

10 μl of Substrate Solution

- 3. Set up a blank control as outlined in step 2, substituting 10 μ l of cold water (4 °C) for the Substrate Solution.
- 4. Initiate each reaction with the addition of 5 μ l of the γ - 32 P-ATP Assay Cocktail, bringing the final reaction volume to 25 μ l. Incubate the mixture in a water bath at 30 °C for 15 minutes.
- 5. After the 15 minute incubation, stop the reaction by spotting 20 μ l of the reaction mixture onto an individually precut strip of phosphocellulose P81 paper.

- Air dry the precut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
- 7. Set up a radioactive control to measure the total γ - 32 P-ATP counts introduced into the reaction. Spot 5 μ l of the γ - 32 P-ATP Assay Cocktail on a precut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
- 8. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
- 9. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

SR = $\frac{\text{cpm of 5} \ \mu \text{l of } \ \gamma^{-32}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$ cpm – value from control (step 7)
nmole – 1.25 nmole (5 μ l of 250 μ M ATP Assay Cocktail)

2. Specific Kinase Activity (SA) (nmole/min/mg)

nmole/min/mg =
$$\Delta$$
cpm × (25/20)
SR × E × T

SR = specific radioactivity of the ATP (cpm/nmole ATP) Δ cpm = cpm of the sample – cpm of the blank (step 3) 25 = total reaction volume

20 = spot volume

T = reaction time (minutes)

E = amount of enzyme (mg)

References

- Beebe, S.J. et al., Molecular cloning of a tissuespecific protein kinase (C gamma) from human testis-representing a third isoform for the catalytic subunit of cAMP-dependent protein kinase. Molec. Endocr., 4, 465-475 (1990).
- 2. Reinton, N. et al., The gene encoding the C gamma catalytic subunit of cAMP-dependent protein kinase is a transcribed retroposon. Genomics, **49**, 290-297 (1998).

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