

TLC Catalog

TLC · Sorbents · Silica Gel · Anhydrous Solvents



2

Introduction to TLC catalog – by Fredric Rabel, Ph.D.



TLC in A World of Chromatography ...

Chromatography is the most widely used analytical technique in the sciences today. It has become an indispensable tool in research, quality control and production. This is particularly true in pharmaceutical, food & beverage, chemistry, and life science arenas. Thin Layer Chromatography (TLC) is just one of a wide range of liquid chromatography techniques used that includes: Liquid Column Chromatography, Solid Phase Extraction (miniaturized version of liquid column chromatography), and High Performance Liquid Chromatography (HPLC). All these techniques utilize the same principle; which states that the speed and efficiency of any separation is controlled by the particle size of the adsorbent and/or the size and configuration of the column or plate.

In the Beginning, there was Merck KGaA ...

In the mid-1950's Merck KGaA, Darmstadt, Germany, was the first company to supply bulk irregular silica for TLC (according to Stahl), and prepared TLC and HPTLC plates (High Performance TLC) on glass, aluminum and plastic supports. These offerings were followed by advances in thin layer technology, resulting in LiChrospher[®] (a spherical silica) and Chromolith[®] (a monolithic silica) TLC Plates which were added to the product line. More recently, Lux TLC plates (which contain a brighter fluorescent indicator for improved detection limits) were made available.

| 1906 | 1938 | | 1949 | 1951-1957 |
|---|---|---|---|---|
| M. Tswett invented chromatography as it is practiced today by causing differential migration of a mixture of chloroplast pigments from a narrow initial zone through a stationary phase in a column by development with a mobile phase. Tswett did not study TLC. | N.A. Izmailov and M.S. Schraiber, at the Pharmaceutical Institute in Kharkov (Ukraine), analyzed plant tinctures by placing a drop of sample solution on a horizontal 2 mm layer of aluminum oxide without binder on a glass microscope slide. Methanol mobile phase was added dropwise to produce | concentric circles of separated zones that were detected by absorption of ultraviolet (UV) light from a lamp. This was the first use of thin layer chromatogram development with mobile phase. It was called "spread layer chromatography" or "spot chromatography." | J.E. Meinhard and N.F. Hall, at the University of Wisconsin, were the first to use binder to prepare layers. They bound a mixture of aluminum oxide and Celite with starch to microscope slides. | J.G. Kirchner and associates, working initially at the USDA laboratories in Pasadena, CA, and then the Coca Cola Company, published methods for TLC on narrow glass "chromatostrips" coated with silicic acid. This is essentially the same technique practiced today. |

The Beauty and Simplicity of TLC ...

Thin Layer Chromatography is perhaps the most under-utilized of all of the chromatography techniques. Although it is plain and simple with no flashing lights or shiny buttons to push, it is surely an economical and efficient technology. There are no large capital investments needed to begin a TLC analysis and most procedures are fairly easy to set up and follow.

Many scientists in Organic and Medicinal Chemistry typically use TLC plates to monitor the progress of organic syntheses. At various points in the process, a sample will be removed to monitor the progress of the synthesis. Alternately, TLC may be used by researchers who are primarily interested in the separation of components. TLC is such a powerful technique that almost any mixture can be separated into its individual components and visualized. TLC can be used to identify or characterize a wide variety of samples and can even be used for preliminary evaluations of samples destined for HPLC analysis. This catalog contains a synopsis of the history of TLC, as well as additional helpful hints that users will find extremely beneficial.

EMD Chemicals is Merck KGaA in North America ...

Always on the forefront of classical and new liquid chromatographic techniques, Merck KGaA, Darmstadt, Germany, continues to produce a full range of silicas and bonded silicas in bulk, columns and on thin layer plates. Contained within this catalog are the highly respected and well trusted products offered by EMD Chemicals.



| 1954 | 1956 | 1958 | 1961 | 1962 |
|---|---|---|--|---|
| R.H. Reitsma, like Kirchner, also used larger glass plates for running several samples side-by-side or 2D TLC and coined the term "chromatoplate". | E. Stahl published his first paper on Dünnschichtchromatogaphie (TLC) and was the first to use the name "thin layer chromatography." | Merck KGaA sold silica gel G (kieselgel G) as well as standardized aluminum oxide and kieselguhr <i>according to Stahl</i> . Stahl was mainly responsible for the standardization of the materials, procedures, and nomenclature of TLC. | Camag began selling TLC equipment such as plate coaters, developing tanks, sample application devices, and UV lamps, and eventually became the leading company in the development of instruments for modern quantitative high performance TLC (HPTLC). | E. Stahl's first TLC book was published containing contributions by a number of TLC specialists; an expanded edition appeared in 1967. Many spray reagents were described in these books. |

Planar Chromatography

The process is an inexpensive and fast separation technique. In its simplest form, it consists of a thin layer of adsorbent on glass, aluminum foil or plastic backing. The adsorbent layer can be any medium. In practice, commercially available products consist predominantly of silica gel and aluminum oxide (in their native or modified forms), cellulose and kieselguhr (diatomaceous earth). The layer thickness generally varies from 0.1 - 0.25 mm for High Performance Thin Layer Chromatography (HPTLC) up to 2 mm thickness for Preparative Thin Layer Chromatography (PTLC).

To determine if an unknown sample consists of a mixture of components, or a known mixture contains all of the intended components, small amounts of sample are transferred as "spots" or "streaks" in a horizontal manner to one side of the adsorbent layer and allowed to dry completely.

The plate is transferred to a "Development Chamber" that contains organic solvent or solvent mixture. Sample spots must remain above the solvent level. Capillary action drives the solvent up the plate over a period of 10 to 20 minutes and separates the sample mixture into its individual components. Occasionally, the correct solvent mixture may have to be determined experimentally. Literature references are a good place to start for developing the protocol that best suits the application.

Adsorbent Layers

Silica gel is by far the most predominant adsorbent layer used in TLC. That may be due to the versatility of silica gel. The typical silica gel used in TLC is an irregularly shaped 15 µm particle with 60Å pores. Also available are TLC plates with adsorbent layers made of irregular and spherical silica particles known as HPTLC plates. These spherical particles are much smaller, close to 5 µm. The advantage of a smaller spherical particle is faster analysis time and lower diffusion of the isolated components. As a result, this medium lends itself to subsequent identification by Raman spectroscopy without removing the components from the plate. A recent development is the monolithic silica gel in which the particles are replaced with a continuous monolithic layer. Separations are even faster than with HPTLC and are completed in a few minutes. The layer is only 10 µm thick and sample capacity is proportionately low.

Other frequently used TLC layers are modified silica gel, notably those modified with hydrocarbons such as dimethyl (RP2), octyl (RP8), and octadecyl (RP18), but also silica gel modifiers such as cyanopropyl, diol, and amine. Less used sorbents are alumina, kieselguhr, cellulose and PEI cellulose. Cellulose's primary area of application is in the life science for the separation of body-fluid extracts, particularly amino acids.

For a detailed treatise on "Sorbents and Precoated Layers in Thin Layer Chromatography," see Handbook of Thin Layer Chromatography, Third Edition, J. Sherma, B. Fried, Marcel Dekker, Inc. 2003.

1968

J.A. Thomas suggested the use of smaller sorbent particles to improve TLC performance (i.e., selectivity, efficiency, and resolution). F. Geiss and H. Schlitt describe the horizontal KS-Vario chamber that allowed testing and optimization of various mobile phases and vapor-saturation conditions. R. Kraffczyk and R. Helger reported laboratory made biphasic plates with a lower 2 cm ion exchange zone for direct desalting of biological samples. H. Halpaap published an important paper on the standardization of commercially precoated plates. A book edited by E.J. Shellard, and a later (1973) book edited by J.C. Touchstone, were very important in promoting the early adoption of quantitative densitometry.

Glass, Aluminum Foil or Plastic-backed Plates

Glass-backed TLC plates are the most commonly used type of plates, but there are many practical reasons for using aluminum or plastic backed TLC plates.

• Aluminum foil and plastic are easy to cut. Instead of using a large 20 x 20 cm plate for just a few samples, a suitable width of the aluminum plate can be cut. The remaining part of the plate can be stored for another analysis.

• Isolated substances can be easily cut from an aluminum or plastic plate, then transferred to an extraction vial for subsequent work up.

Glass is heavy and expensive to transport.

Glass plates take up more storage space.
 Five glass plates take up as much space as a 25 pack of aluminum or plastic plates.

Cutting and Breaking Analytical Glass TLC Plates

Although a wide selection of prescored TLC plates is available, some researchers choose to use large 20 x 20 cm TLC plates exclusively. If the need arises, a large TLC plate can be cut to a smaller size. Depending on the specific application, one large plate can be cut to eighty (80) 1 x 2.5 cm plates size or 2,000 microscope slide plates from a box of 25.

Great care must be taken to avoid injury or waste when cutting glass TLC plates. Wear protective gear including gloves and safety glasses. Lay the glass plate, sorbent side down, onto a large sheet of filter paper on a clean, smooth, flat surface. Outline the dimensions with a suitable marker and a straight edge. Holding the straight edge where the initial cut is to be made, cut with a scoring device. (Special multiple-score cutting devices are available in craft, stained glass, or paint stores.) Next, snap along the score line with a quick pull and bend. Use of a grozier (a snapping tool much like pliers available at the above mentioned stores) is recommended.

Precut and Prescored TLC Plates

In a commercial laboratory setting, productivity is the overriding theme. To this end EMD Chemicals offers a series of different sizes of precut plates. Precut TLC plates are available in sizes of 2.5 x 7.5 cm, 5 x 10 cm, 5 x 20 cm, 10 x 10 cm, and 10 x 20 cm. Similarly, a series of prescored glass plates are commercially available as 2.5 x 10 cm, 5 x 10 cm, and 5 x 20 cm.

Precut plates are convenient and easy to use, plus prescored plates add safety and versatility to any laboratory. The plates can be used "as is" or broken down to the required size without loss of continuity in the adsorbent layer.



1968-1971

1970

The earliest commercial densitometers for direct quantitative TLC were manufactured by Zeiss, Kontes, Camag, Turner, Schoeffel, and Vitatron. The first silica gel 60 TLC plates silanized with RP2 were produced by Merck KGaA. D. Jaworek reported the first use of laboratory made Sephadex layers for the size exclusion TLC of proteins for molecular weight measurement. J. Sherma initiated biennial reviews of the literature of TLC for the Fundamental Reviews issues of the American Chemical Society journal Analytical Chemistry that have continued to be published continually through 2004.

1971

LR. Snyder described a systematic theory of adsorption chromatography involving the concepts of selectivity, resolution, and theoretical plates. Several years later he introduced the selectivity triangle, the use of which aided the choice of optimum mobile phases for TLC.

TLC or HPTLC

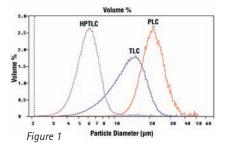
The predominant medium for TLC is silica gel 60, which is a silica gel particle that has a 60 Å mean pore size. In TLC, separation time and efficiency are functions of particle size and particle size distribution. In general, the mean particle size of standard TLC plates is 15 μ m, while that for HPTLC plates is ~5 μ m (see Fig. 1). The layer thickness for standard TLC plates is 0.25 mm (250 μ m) and is 0.2 mm for HPTLC. Advantages to using HPTLC plates are:

Faster analysis

processes of TLC.

- Higher efficiency
- · Lower diffusion during sample migration
- · Improved detection limit

In contrast to HPTLC, standard TLC plates have larger sample capacity. Sometimes this larger capacity allows for the use of standard TLC plates for isolation and recovery in place of the larger and more expensive preparative plates.



Polymer Bonded (Hard-Layer or Soft-Layer) Gypsum Binder TLC Plates

During the early development stages of TLC, gypsum (Plaster of Paris) was added in a 10 to 20% concentration to silica gel to bind the slurry to the glass plate. Early practitioners used this process to prepare their own plates. This bond, formed between the layer and the substrate, was very weak and chipped easily. Preserving the silica gel plates prepared with gypsum binder was difficult and often required additional precautionary steps to prevent the loss of the surface.

When TLC plates became commercially available, it became quickly apparent that a stronger binder was needed. There are a number of binders that can be used. TLC plates from EMD Chemicals are produced with 1% acrylate as a binder. This concentration is sufficient to bind the layer to the plate during shipping, but not so high as to interfere with the separation process. The two advantages often cited for using "G" plates (plates with gypsum binder) are the ease of removing isolated fractions from the "soft" layer, and the ability of being able to char the plate with sulfuric acid. In practice these advantages are somewhat overstated. Isolated substances are easily recovered from plates containing an organic binder and "careful" charring with sulfuric acid makes it possible to differentiate between the darkened plate and the charred spots.

The big disadvantage of the softer "G" plates is that the adsorbent layer is very fragile and fractures easily. This makes it almost impossible to cut the plates into smaller sizes.

Many TLC methods developed with gypsum (G) binder plates are still referenced. The cited plates can be easily substituted with the newer polymer bound plates and often give better results. USP/NF allows gypsum, starch or other binders in the TLC plates used in their protocols.

| 1972 | 1975 | | 1978 | 1979 |
|---|---|--|---|--|
| A book by F. Geiss was published that presented the theoretical and procedural aspects of TLC in detail. This book became the definitive guideline for performing TLC and is still widely used today to gain an understanding of the underlying | F. Eisenbeiss prepared a layer with 5 μm diameter silica gel particles for high performance TLC (HPTLC) in 1975, and commercial silica gel 60 HPTLC plates were first presented by Merck KGaA about this time. | J.A. Perry, T.H. Jupille, and L.J. Glunz introduced programmed multiple development, a method for layer development with a single solvent over increasing distances. | Merck KGaA introduced RP2, RP8 and RP18 phases as precoated HPTLC plates. | E. Tyihak, E. Mincsovics, and H. Kalasz described overpressured layer chromatography (OPLC) for fast separations with constant mobile phase flow and theoretical plate height over the complete layer distance. |

TLC Plates with Sample Concentration Zone

The concentration zone layer consists of a 2.5 cm band of wide-pore silica gel particles along one edge of the plate. The large pore size of this part of the layer offers no noticeable resistance to sample migration when the plate is developed. As a result, when the TLC plate is spotted with the sample anywhere in the concentration zone, it quickly moves as a sample band to the active silica gel portion of the plate where separation begins.

Using plates (with a pre-concentration zone) provide several advantages:

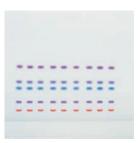
 \cdot Eliminates aligning samples at the origin.

 \cdot Sample fractions and corresponding standards are sure to migrate at the same speed.

 \cdot Excessive sample amounts will result in the same separation efficiency.







Water Tolerant TLC/HPLC Plates

The organic binder used in the prepared TLC and HPTLC plate is an acrylate. This is a water soluble binder which can swell the layers when used in developing solvents with high water content (generally > 50%). This is not often seen with silica gel layers, but can occur more often with bonded layer TLC plates, like a RP-18 plate.

To allow the chromatographer to get better results, Merck KGaA has reformulated some plates to be used with high water content mobile phases for both Silica Gel and RP-18 plates. These products can be found throughout the following product listing by reading the heading or comments beside each product. The letter "W" is used in the product description of these products.

| 1979 | 1980 | 1981 | 1984 | |
|---|--|---|---|--|
| Camag manufactured the first monochromator-equipped densitometer, the TLC/HPTLC Scanner I. | F. Kreuzig described a mechanized spraying device for application of derivatization reagents to be used for zone detection. | RP8 and RP18 plates were offered as precoated TLC plates by Merck KGaA. | K. Burger introduced automated multiple development (AMD) for development of layers over increasing distances with a stepwise mobile phase gradient of decreasing strength. | The first successful experiments with digital cameras for the quantitative evaluation of chromatograms were reported by M. Prosek, A. Medija, M. Katic, and R.E. Kaiser. |

Preparing the TLC Plate For Use

There are two general preparation steps for TLC plates: washing and activation. It is recommended that you wash the plates before the sample is applied with the same solvent intended for the separation. This removes any substance that may have adsorbed to the plate during storage. The plate can be either totally immersed in the solvent or the solvent can be allowed to migrate up the plate in a development chamber overnight.

Activating a glass backed silica gel plate is highly recommended because it improves reproducibility of your results. This is especially important for repetitive analysis when results are compared across a span of days or weeks. Heat the glass TLC plate in a hot-air oven for 30 minutes at 110°C. Then remove the plate from the oven and immediately place it into a humidity controlled environment, such as a storage box or desiccator. Plates left out in the open will absorb moisture quickly, reversing the activation.

Activation of TLC plates on aluminum or plastic backing should be done at 90°C and should be performed on a solid metal or glass surface to ensure uniform heat distribution.

1991

The Benefits of TLC ...

There are significant benefits in using TLC as an accessory tool to HPLC or in screening tens to hundreds of samples in a single day in high throughput applications.

 \cdot Substances that remain on the origin are easily identified and recovered, resulting in appropriate conditions to effect separation without losing the HPLC column or the sample in question.

• The type and number of solvents available for TLC chromatography has expanded to include those without UV transparency, since the solvent is evaporated before the components are elucidated.

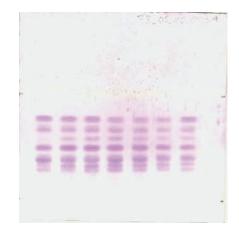
• Detection methods other than spectrometric and spectrophotometric methods can be used to identify isolated substances.

 \cdot As many as 70 samples and reference standards can be chromatographed on a single standard TLC Plate.

• Solvent consumption is minimal and sometimes as low as 10 ml for as many as 70 samples. • Two-dimensional or multi-dimensional separations on the same TLC plate is possible by simply allowing the solvent from the first development step to evaporate, turning the plate 90 degrees, and then using a different solvent mixture to continue the separation process.

• Coupling TLC with other separation techniques or spectrophotometric methods is readily accomplished because fractions are easily recovered from the plates for subsequent analysis.

The EMD advantage: we produce our own silica gel and therefore have complete control over its purity.



1988

Analtech produced the first commercial video densitometer in 1988.

J. Sherma and B. Fried edited for Marcel Dekker Inc. the first of three editions of the Handbook of Thin Layer Chromatography. These are the only books ever published that cover in detail essentially all of the important knowledge of TLC theory, techniques, instrumentation, and applications to virtually every analyte and sample type in more than 30 chapters and 1,000 pages.

Seeing the difference ... Visualization Techniques

Fluorescence Quenching – Like any separation technique, the key is to find ways to visualize or quantify the results of the separation. For TLC, one of the best ways to visualize the separation is to use the fluorescent indicator already contained in the plate (TLC Plates designated F_{254} contain a fluorescent indicator). This technique, called fluorescence quenching, results in the dark spots of the separated sample components showing up clearly against a fluorescent background.

Although use of TLC plates with fluorescence background is the most popular method to visualize isolated fractions, plates can also be impregnated with other reagents, dried and stored for future use. A few common reagents are ammonium sulfate in place of sulfuric acid, magnesium acetate, potassium oxalate and silver nitrate. **Reagent Visualization** – Sometimes a special visualizer solution needs to be sprayed over the developed plate in order to form visible spots. One example would be the use of a Ninhydrin solution to visualize the resulting spots formed by the separation of amino acids.

There are hundreds of potential visualization reagents and their selection depends on the molecular structure and functional groups of the sample being studied. Refer to the following publications for additional information.

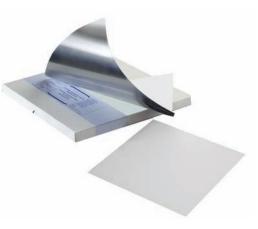
Spot Tests in Organic Chemistry, 4th reproduction, F. Fiegl & V. Anger (Elsevier, NY, 1989)

Thin Layer Chromotography, 2nd edition, Egon Stahl, editor (Springer-Verlag, NY, 1969)

TLC Reagents & Detection Methods, Vol. 1a, H. Jork & W. Funk, et al., (Wiley, NY, 1990)

EMD Chemicals has produced a TLC Visualization Guide with over 300 examples. It is now available on our website (www. emdchemicals.com/analytics) for easier accessibility and convenience. Select the TLC Visualization Reagents guide pdf and search for the type of samples you need to visualize.

Charring - One of the oldest ways to identify isolated substances on a TLC plate is by charring with sulfuric acid. After the plate has been developed and residual solvent is completely removed, it is placed in a glass housing or other protected container in a fume hood and sprayed with a fine mist of diluted sulfuric acid. The plate is subsequently placed in an oven at 150°C or on a hot plate to allow the sulfuric acid to decompose the sample spots. When using plates that contain an organic binder, lower heat is applied so as not to char the plate along with the sample spots. Optional spraying reagents are ammonium sulfate and phosphoric or phosphomolybdic acids.



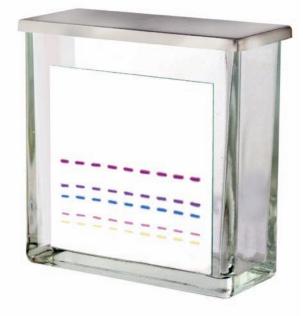
Reference:

Joseph Sherma, Thin Layer Chromatography, in A Century of Separation Science, H.J. Issaq (editor), Marcel Dekker Inc., New York, NY, 2002, pp. 49-68. Joseph Sherma Department of Chemistry Lafayette College Easton, PA 18042

Some important TLC considerations ...

Although it is not difficult to get good TLC results, the reproducibility of results is paramount. For highly consistent and reliable results, start by selecting a TLC plate from EMD Chemicals. To get started, it is recommended to always heat activate the plate before spotting it with the sample. Then, when the plate is ready to be placed into the development chamber, make sure that the chamber is lined with filter paper and has been sufficiently equilibrated with the solvent mixture.

For faster results and improved detection limits, choose the HPTLC plate that is equivalent to the TLC Plate being used. The HPTLC plates are coated with smaller particles which results in shorter development distances. TLC and HPTLC plates that have a sample concentration zone will always provide cleaner separations by converting the sample spot into a sample band. This sample band lines up the sample components into a more uniform alignment so they all begin their migration together instead of in a random staggered configuration. For additional technical assistance for developing your TLC methodology, please visit our web site (www.emdchemicals.com) or call us at 800 222-0342.



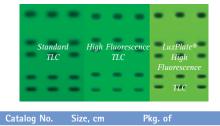
New Products

LuxPlate[®] High Fluorescence TLC Plates

LuxPlate TLC Plates contain a high concentration of fluorescence indicator which has major advantages over standard TLC plates. The new LuxPlate TLC Plates provide:

- \cdot Twice the brightness
- \cdot Improved visualization of fraction
- \cdot Higher binder content to increase layer stability
- \cdot Same separation performance as standard plates

LuxPlate TLC Plates are available in several different plate sizes including the popular 2.5 x 7.5 cm plate.



With F_{254} indicator

| 204 | | | |
|--------|-----------|-----|------|
| 5801-1 | 2.5 x 7.5 | 100 | SI60 |
| 5802-1 | 5 x 10 | 25 | SI60 |
| 5803-1 | 5 x 20 | 100 | SI60 |
| 5804-1 | 10 x 20 | 50 | SI60 |
| 5805-1 | 20 x 20 | 25 | SI60 |

Ultra-thin, Monolithic-layer TLC Plates

These plates are the latest in a long line of innovations in Planar Chromatography. Similar to Chromolith columns, the UTLC plates consists of a monolithic, 0.01 mm = 10 μ m layer of Silica gel. The layer consists of particles with pore diameters of 30 to 40 Å and 1 to 2 μ m, respectively. Advantages are:

- \cdot Nanograms of samples can be isolated and identified
- \cdot Migration distance is 1 to 3 cm
- \cdot Analysis time is 1 to 6 minutes

5007-1 25/pkg.

Physical Properties

| Monolithic Layer | No particles |
|-----------------------|------------------------------|
| Plate Dimensions | 60 mm X 36 mm |
| Layer Thickness | 10 µm |
| Meso Pores | 30 to 40 Å pore diameter |
| Macro Pores | 1 to 2 μm pore diameter |
| Specific surface area | ~350 m²/g |
| Specific pore volume | ~0.3 ml/g (meso Pores) |

The medium coating these unique UTLC (Ultra-Thin TLC) plates is actually a porous layer of silica gel based on patented technology. These UTLC plates offer several key advantages over conventional and other high performance plates including:

- · Dramatically shortened development time
- Elimination of any binder because the gel adheres to the plate by itself
- Very low solvent consumption
- · High separation efficiency
- · High analytical sensitivity
- $\cdot \,$ Stability in water



Photo showing actual monolithic gel coating structure.

TLC Glass Plates

October Nie

Silica Gel 60 on Glass Plates - 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|-----------|---------|--------------------------|
| | | | |
| 15326-1 | 2.5 x 7.5 | 100 | Includes Box for storage |
| 105639-5 | 2.5 x 7.5 | 500 | |
| 5724-3 | 5 x 20 | 100 | |
| 5626-6 | 10 x 20 | 50 | |
| 5721-7 | 20 x 20 | 25 | |
| With F ₂₅₄ indicator | | | |
| 15327-1 | 2.5 x 7.5 | 100 | Includes Box for storage |
| 15341-1 | 2.5 x 7.5 | 100 | |
| 15341-5 | 2.5 x 7.5 | 500 | |
| 5789-2 | 5 x 10 | 25 | |
| 5719-2 | 5 x 10 | 200 | |
| 5808-3 | 5 x 20 | 25 | |
| 5714-3 | 5 x 20 | 100 | |
| 5729-6 | 10 x 20 | 50 | |
| 5715-7 | 20 x 20 | 25 | |
| | | | |

Silica Gel 60 with F₂₅₄ indicator on Glass Plates - 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---|
| PRE-SCORED | | |
| 5608-7 | 20 x 20 | 20 Prescored to 5 x 20 cm segments |
| 10557-1 | 20 x 10 | 25 Prescored to 5 x 20; 2.5 x 10; 5 x 20 or 5 x 10 cm segments |
| 105620-7 | 20 x 20 | 25 Prescored to 5 x 10 cm segments |

Silica Gel 60 with F_{254} indicator on Laser coded Glass Plates – 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|-----------------------------|
| Laser Coded | | |
| | | |
| 105566-7 | 20 x 20 | 25 Laser coded for tracking |

Silica Gel 60 on Glass Plates - 0.25 mm coating; with 2.5 cm Concentration zone

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|----------|---------|--|
| | | | |
| 11844-6 | 10 x 20 | 50 | |
| 11845-7 | 20 x 20 | 25 | |
| With F ₂₅₄ indicator | | | |
| 11846-6 | 10 x 20 | 50 | |
| 11798-7 | 20 x 20 | 25 | |



TLC Glass Plates

Reversed Phase (RP) Silica Gel 60 on Glass Plates - 0.25 mm Coating

| Catalog No. | Bonded Phase | Size, cm | Pkg. of |
|----------------------------------|--------------|----------|---------|
| 5746-7 | RP-2 | 20 x 20 | 25 |
| With F ₂₅₄ indicator | | | |
| 5747-7 | RP-2 | 20 x 20 | 25 |
| With F _{254s} indicator | | | |
| 15684-1 | RP-8 | 5 x 10 | 25 |
| 15682-3 | RP-8 | 5 x 20 | 50 |
| 15424-6 | RP-8 | 10 x 20 | 50 |
| 15388-7 | RP-8 | 20 x 20 | 25 |
| 15685-1 | RP-18 W | 5 x 10 | 25 |
| 15683-3 | RP-18 W | 5 x 20 | 50 |
| 15423-6 | RP-18 W | 10 x 20 | 50 |
| 15389-7 | RP-18 W | 20 x 20 | 25 |

| Silica Gel 60 W with F254c indic | ator on Glass Plates - | Water Resistant 0.20 mm Coating | |
|----------------------------------|------------------------|---------------------------------|--|
| 2545 | | ······ | |

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| | | |
| 16485-1 | 20 x 20 | 25 |

Silica Gel 40 with F₂₅₄ indicator on Glass Plates - 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| | | |
| 5634-7 | 20 x 20 | 25 |

Kieselguhr with F_{254} indicator on Glass Plates – 0.2 mm Coating

(Silica Gel/Kieselguhr/Diatomaceous Earth)

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|-------------------------------|
| | | |
| 5738-7 | 20 x 20 | 25 Kieselguhr |
| 5737-7 | 20 x 20 | 25 Silica Gel 60 - Kieselguhr |

Aluminum Oxide 60 (Basic) with F_{254} indicator on Glass Plates – 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| 5701 0 | 5 20 | 100 |
| 5731-3 | 5 x 20 | 100 |
| 5713-7 | 20 x 20 | 25 |

Aluminum Oxide 150 (Basic) with F₂₅₄ indicator on Glass Plates - 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| 5707 7 | 20 20 | 25 |
| 5727-7 | 20 x 20 | 25 |

Cellulose on Glass Plates - 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| | | |
| 5632-5 | 10 x 10 | 100 |
| 5730-6 | 10 x 20 | 50 |
| 5716-7 | 20 x 20 | 25 |

Cellulose F on Glass Plates - 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| | | |
| 5728-6 | 10 x 20 | 50 |
| 5718-7 | 20 x 20 | 25 |

PEI Cellulose F on Glass Plates - 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|-----------------|
| 5725-7 | 20 x 20 | 25 Store at 4°C |

TLC Aluminum Sheets

Silica Gel 60 on Aluminum Sheets - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|--------------------------------|----------|---------|--|
| | | | |
| 16835-2 | 5 x 10 | 50 | |
| 5553-7 | 20 x 20 | 25 | |
| With F ₂₅₄ indicato | r | | |
| 5549-4 | 5 x 7.5 | 20 | |
| 16834-2 | 5 x 10 | 50 | |
| 5534-3 | 5 x 20 | 100 | |
| 5554-7 | 20 x 20 | 25 | |
| 5562-7 | 500 x 20 | 1 Roll | |
| | | | |

Silica Gel 60 on Aluminum Sheets - 0.2 mm Coating with 2.5 cm Preconcentration Zone

| | | 5 |
|---------------------------------|----------|---------|
| Catalog No. | Size, cm | Pkg. of |
| | | |
| 5582-7 | 20 x 20 | 25 |
| With F ₂₅₄ indicator | | |
| 5583-7 | 20 x 20 | 25 |
| | | |

Silica Gel 60 W on Aluminum Sheets - Water Resistant 0.20 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|---------------------------------|----------|---------|
| | | |
| 16487-1 | 20 x 20 | 25 |
| With F ₂₅₄ indicator | | |
| 16484-1 | 20 x 20 | 25 |

Kieselguhr with F_{254} indicator on Aluminum Sheets – 0.2 mm Coating

(Silica Gel/Kieselguhr/Diatomaceous Earth)

| Catalog No. | Size, cm | Pkg. of | |
|-------------|----------|---------|----------------------------|
| | | | |
| 5568-7 | 20 x 20 | 25 | Kieselguhr |
| 5567-7 | 20 x 20 | 25 | Silica Gel 60 |
| 105533-7 | 20 x 20 | 20 | Silica Gel NH ₂ |

Aluminum Oxide 60 (Neutral) with F₂₅₄ indicator Aluminum Sheets - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| | | |
| 5550-7 | 20 x 20 | 25 |

Aluminum Oxide 150 (Neutral) with F_{254} indicator Aluminum Sheets - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|-------------|----------|---------|
| | | 25 |
| 5551-7 | 20 x 20 | 25 |

Cellulose on Aluminum Sheets - 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|----------|---------|--|
| | | | |
| 5552-7 | 20 x 20 | 25 | |
| 5563-7 | 500 x 20 | 1 Roll | |
| With F ₂₅₄ indicator | | | |
| 5574-7 | 20 x 20 | 25 | |

TLC Plastic Sheets

Silica Gel 60 on Plastic Sheets - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|----------|---------|--|
| | | | |
| 5748-7 | 20 x 20 | 25 | |
| With F ₂₅₄ indicator | r | | |
| 5735-7 | 20 x 20 | 25 | |
| 5749-7 | 500 x 20 | 1 Roll | |

Aluminum Oxide 60 (Neutral) with F_{254} indicator on Plastic Sheets – 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|-------------|----------|---------|--|
| 5581-7 | 20 x 20 | 25 | |
| 00017 | 20 X 20 | 20 | |

Cellulose on Plastic Sheets - 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|----------|---------|--|
| | | | |
| 5577-7 | 20 x 20 | 25 | |
| With F ₂₅₄ indicator | | | |
| 5565-7 | 20 x 20 | 25 | |

Letter and Number Definitions

| AMD | Automated multiple development after Camag |
|-----------------------|---|
| CHIR | Chiral layer for separating enantiomers of amino acids |
| CHIR | Chiral layer for separating enantiomers of amino acids |
| CN | Hydrophilic layer with cyano modification |
| DIOL | Hydrophilic layer with diol modification |
| F | Containing fluorescent indicator |
| H (bulk sizes) | Containing no foreign binders |
| HR (bulk sizes) | Specially purified |
| NH2 | Hydrophilic layer with amino modification |
| P | For preparative layers |
| PEI | Polyethyleneimine |
| PLC | Preparative layer chromatography |
| R | Specially purified |
| Silanized RP-2 | Reversed phase with a dimethylsilyl chain |
| RP-18 | Reversed phase with a C18 hydrocarbon chain |
| Silica Gel 60Å | Silica gel with a mean pore size of 60 Angstrom |
| TLC | Thin layer chromatography |
| W | Water tolerant wettable layer |
| 40, 60, etc. | Mean pore size in Å |
| Pressored | Plate can be broken down into smaller plates |
| Preconcentration zone | An area of the TLC plate composed of silica gel of large, 50,000 Å, pore size |

PEI Cellulose with F_{254} indicator on Plastic Sheets – 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|-------------|----------|---------|--------------|
| 5570 7 | 20 20 | 25 | Share at 490 |
| 5579-7 | 20 x 20 | 25 | Store at 4°C |

Preparative TLC Glass Plates

PTLC Silica Gel 60 on Glass Plates

| Catalog No. | Size, cm | Pkg. of | Indicator | Layer Thickness |
|-------------|----------|---------|-----------------------------------|-----------------|
| | | | | |
| 13894-7 | 20 x 20 | 20 | None | 0.5mm |
| 5744-7 | 20 x 20 | 20 | F ₂₅₄ | 0.5mm |
| 13895-7 | 20 x 20 | 15 | F ₂₅₄ | 1 mm |
| 5745-7 | 20 x 20 | 12 | None | 2mm |
| 5717-7 | 20 x 20 | 12 | F ₂₅₄ | 2mm |
| 5637-7 | 20 x 20 | 12 | F ₂₅₄ + ₃₆₆ | 2mm |

PTLC Silica Gel 60 on Glass Plates - 1 mm Water Resistant Coating Chemically Modified Layers

| Catalog No. | Bonded Phase | Size, cm | Pkg. of | Indicator | |
|-------------|--------------|----------|---------|-------------------|--|
| 5434-7 | RP-18 W | 20 x 20 | 15 | F _{254s} | |

PTLC Silica Gel 60 with F_{254} indicator on Glass Plates With 4 x 20 cm Preconcentration Zone

| Catalog No. | Size, cm | Pkg. of | Layer Thickness | |
|-------------|----------|---------|-----------------|--|
| | | | | |
| 13792-7 | 20 x 20 | 15 | 1 mm | |
| 13793-7 | 20 x 20 | 12 | 2mm | |
| 13794-7 | 20 x 20 | 20 | 0.5mm | |

PTLC Aluminum Oxide 60 with F₂₅₄ indicator on Glass Plates - 1.5 mm Coating

20 x 20

| Catalog No. | Size, cm | Pkg. of | - |
|-----------------|-----------------------------------|---|----------|
| 5788-7 | 20 x 20 | 12 | <i>H</i> |
| PTLC Aluminum C | Dxide 150 with F ₂₅₄ i | ndicator on Glass Plates - 1.5 mm Coating | |
| Catalog No. | Size, cm | Pkg. of | |

12

Fluorescent Indicator

9182-1 50g bottle of F₂₅₄ Fluorescent Indicator, for use with any TLC plates

HPTLC/TLC Devices - UV Light

The portable UV lights are only $1\times3.5\times6"$ in size and are battery operated (Batteries not included, requires 5 "C" size). They are easily taken into the field or used in the laboratory.

| 13203-1 | Portable UV Light, 366 nm | 1 Piece, complete |
|---------|---------------------------|-------------------|
| 12537-1 | Portable UV Light, 254 nm | 1 Piece, complete |

TLC Book

This book on thin layer chromatography gives many of the essentials of the technique. It is ideal for the beginner or for those people needing a general review of the various techniques used today in TLC. (66 pages)

| 1000-1 | Thin Layer Chromatography: An Introduction, K. Bauer, L. Gross and W. Sauer |
|--------|---|
| | Merck KGaA, Darmstadt, Germany, 1990 |

TLC Learning Program

An attractively priced computer-based interactive learning program for TLC and HPTLC.

| 960314-1 | Modern Thin Layer Chromatography (3.5" diskettes, 1.44MB, only; for PC, IBM, XT, |
|----------|--|
| | AT, PS/2 or compatible, 8MB required for installation, 572KB RAM, EGA, VGA or |
| | compatible, MS-DOS, 3.3 and above) |

5726-7

HPTLC Glass Plates

HPTLC Silica Gel 60 on Glass Plates - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|----------------------------------|----------|---------|--|
| | | | |
| 5631-5 | 10 x 10 | 25 | |
| 5633-5 | 10 x 10 | 100 | |
| 5641-6 | 10 x 20 | 50 | |
| 116436-1 | 20 x 20 | 25 | |
| With F ₂₅₄ indicator | - | | |
| 105616-2 | 5 x 10 | 25 | |
| 5628-5 | 10 x 10 | 25 | |
| 5629-5 | 10 x 10 | 100 | |
| 5642-6 | 20 x 10 | 50 | |
| 115534-1 | 20 x 20 | 25 | |
| With F _{254s} indicator | r | | |
| 15696-6 | 10 x 20 | 25 | |

HPTLC Silica Gel 60 on Glass Plates - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|----------|---------|--------------------------------|
| | | | |
| Prescored | | | |
| 5644-5 | 10 x 10 | 25 | Prescored to 5 x 5 cm segments |
| With F ₂₅₄ indicator | | | |
| 5635-5 | 10 x 10 | 25 | Prescored to 5 x 5 cm segments |

HPTLC Silica Gel 60 with F₂₅₄ indicator on Glass Plates - 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|-------------|----------|---------|--------------------------|
| Laser Coded | | | |
| 105564-5 | 10 x 10 | 25 | Laser coded for tracking |
| 5613-6 | 10 x 20 | 25 | Laser coded for tracking |

HPTLC Silica Gel 60 W with F_{254s} indicator on Glass Plates - Water Resistant Coating

| Catalog No. | Size, cm | Pkg. of | |
|--------------|----------|--|-----------|
| 1.05646.0001 | 20 x 10 | 25 Lichrospher® 6-8 mm p RP18 bonded 0.2 mm c | |
| 1.12363.0001 | 20 x 20 | 25 Suitable for AMD 0.1 m | m coating |

HPTLC Silica Gel 60 on Glass Plates - 0.2 mm Coating with 2.5 cm Concentration Zone

| Catalog No. | Size, cm | Pkg. of |
|---------------------------------|----------|---------|
| | | |
| 13748-5 | 10 x 10 | 25 |
| 13749-6 | 20 x 10 | 50 |
| With F ₂₅₄ indicator | | |
| 13187-1 | 5 x 10 | 25 |
| 13727-5 | 10 x 10 | 25 |
| 13728-6 | 20 x 10 | 50 |

HPTLC Glass Plates

HPTLC Silica Gel 60 on Glass Plates - 0.25 mm Coating

| Catalog No. | Size, cm | Pkg. of | Bonded Phase | Fluorescent Indicator |
|-------------|----------|---------|-----------------|-----------------------|
| | | | | |
| 14101-1 | 10 x 10 | 25 | CHIR | None |
| 16464-5 | 10 x 10 | 25 | CN | F _{254s} |
| 12571-6 | 10 x 20 | 25 | CN | F _{254s} |
| 12668-5 | 10 x 10 | 25 | DIOL | F _{254s} |
| 12572-6 | 10 x 20 | 25 | NH ₂ | None |
| 15647-5 | 10 x 10 | 25 | NH ₂ | F _{254s} |
| 13726-5 | 10 x 10 | 25 | RP-2 | F _{254s} |
| 13725-5 | 10 x 10 | 25 | RP-8 | F _{254s} |
| 5914-6 | 10 x 20 | 25 | RP-18 | None |
| 13724-5 | 10 x 10 | 25 | RP-18 | F _{254s} |
| 15498-6 | 20 x 10 | 25 | RP-18 | F _{254s} |

Specialty HPTLC Silica Gel 60 on Glass Plates

| Catalog No. | Size, cm | Pkg. of | Description |
|--------------|-----------|---------|--|
| | | | |
| 15552-6 | 10 x 20 | 25 | F _{254s} indicator with Water Resistant 0.2 mm Coating - for AMD |
| 1.12363.0001 | 20 x 10 | 25 | F _{254s} indicator with Water Resistant 0.1 mm Coating - for AMD |
| 11764-6 | 10 x 20 | 25 | $\rm F_{254s}$ indicator with 0.1 mm Coating - for AMD |
| 5787-5 | 10 x 10 | 25 | No indicator with 0.1 mm Coating |
| 15036-6 | 10 x 20 | 50 | F254s indicator Cellulose - 0.1 mm Coating |
| 115445-1 | 20 x 10 | 25 | F254s indicator on LiChrospher® HPTLC |
| 5007-1 | 60 x 36mm | 25 | No indicator - Ultra Thin Plate, Chromolith Silica UTLC |
| 5613-6 | 20 x 10 | 25 | $\mathrm{F}_{\mathrm{254}}$ indicator for GLP with 2 mm coating GLP HPTLC |

HPTLC Silica Gel 60 with Chemically Modified Layers on Glass Plates - 0.2 mm Coating

With 2.5 cm Pre-Concentration Zone

| Catalog No. | Bonded Phase | Size, cm | Pkg. of | ; |
|-------------|--------------|----------|---------|-------------------------------------|
| | | | | |
| 15037-6 | RP-18 | 20 x 10 | 25 | For PAH detection in drinking water |

HPTLC Silica Gel 60 with Chemically Modified Layers on Glass Plates

0.2 mm Water Resistant Coating

| Catalog No. | Bonded Phase | Size, cm | Pkg. of | Fluorescent Indicator |
|-------------|--------------|----------|---------|-----------------------|
| | | | | |
| 14296-6 | RP-18 W | 10 x 20 | 25 | None |
| 13124-1 | RP-18 W | 10 x 10 | 25 | F _{254s} |

HPTLC Aluminum Sheets

HPTLC Silica Gel 60 on Aluminum Sheets - 0.2 mm Coating

| Catalog No. | Size, cm | Pkg. of |
|--------------------------------|----------|---------|
| | | |
| 5547-7 | 20 x 20 | 25 |
| With F ₂₅₄ indicate | or | |
| 5556-4 | 5 x 7.5 | 20 |
| 5548-7 | 20 x 20 | 25 |

HPTLC Silica Gel 60 with F_{254s} indicator on Aluminum Sheets - 0.2 mm Coating

| | 2015 | | |
|----------------------------|-------------|----------|---------|
| Bonded Phase | Catalog No. | Size, cm | Pkg. of |
| | | | |
| Chemically Modified | Layers | | |
| DD 10 | 5500 4 | F 7 F | 20 |

| RP-18 | 5560-4 | 5 x 7.5 | 20 |
|-------|--------|---------|----|
| RP-18 | 5559-7 | 20 x 20 | 20 |

Specialty HPTLC Silica Gel 60 on Aluminum Sheets

| Catalog No. | Size, cm | Pkg. of | Indicator | Specialty |
|-------------|----------|---------|-------------------|--|
| 105543-1 | 10 x 10 | 25 | F _{254s} | Raman 6-8µm - 0.1 mm Coating |
| 105586-1 | 20 x 20 | 25 | F _{254s} | LiChrospher® 3-5 µm - 0.1mm Coating |

HPTLC Cellulose on Aluminum Sheets - 0.1 mm Coating

| Catalog No. | Size, cm | Pkg. of | |
|---------------------------------|----------|---------|--|
| | | | |
| 5786-6 | 10 x 20 | 50 | |
| 16092-1 | 20 x 20 | 25 | |
| With F ₂₅₄ indicator | | | |
| 15035-5 | 10 x 10 | 25 | |



DriSolv[®] Anhydrous Solvents

EMD Chemicals is a recognized leader in the production of high purity solvents. We use that experience to produce the highest quality anhydrous solvents. With a water content as low as 10 ppm for some solvents, our DriSolv[®] solvents provide a reliable basis for water critical synthesis and research work.

All DriSolv® solvents are packed under nitrogen and sealed with our Pureseal® septa, which ensures that the solvent will stay dry during use. These unique septa have been designed to reseal after every use and are chemically inert. The large surface area enables access for more than one cannula and multiple reclosing options, making it easy to deliver the driest solvents to any type of reaction vessel.

In addition to the standard 100 ml and 1L bottles for laboratory use, our DriSolv[®] solvents are available in stainless steel EM ReCycler[®] containers in sizes from 18.9L to 1,250L.



Acetonitrile, DriSolv®

| Methyl cyanide | | |
|--------------------|----------------|------------------|
| CH ₃ CN | | |
| Specifications: | | |
| Appearance | clear, free fi | rom particulates |
| Assay (GC), % | | ≥ 99.8 |
| Color (APHA) | | ≤ 10 |
| Evaporation resid | ue, ppm | ≤ 1 |
| Water, ppm | | ≤ 50 |
| ltem | Size | |
| AX0143-6 | 1L | |
| | 6 x 1L | |
| AX0143-7 | 100 ml | |
| | 6 x 100 ml | |
| | | |

Alcohol, Reagent, DriSolv®

| СН ₃ СН ₂ ОН | |
|------------------------------------|-------------------------------|
| Specifications: | |
| Appearance | clear, free from particulates |
| Assay (GC), % | Ethanol 89.5 - 91.5% |
| | Methanol 3.5 - |
| 5.5% | |
| | lsopropanol 4.0 - 6.0% |
| Color (APHA) | ≤ 10 |
| Residue after evaporat | tion, ppm ≤ 5 |
| Water, ppm | |
| | C 1 |
| ltem | Size |
| AX0447-6 | 1L |
| | 6 x 1L |
| AX0447-7 | 100 ml |
| | 6 x 1L |
| | |

Benzene, DriSolv®

| с ₆ н ₆ | |
|-------------------------------|-------------------------------|
| Specifications: | |
| Appearance | clear, free from particulates |
| Assay (GC), % | ≥ 99.8 |
| Color (APHA) | ≤ 10 |
| Evaporation residue | , ppm ≤ 5 |
| Water, ppm | ≤ 30 |
| | <u> </u> |
| ltem | Size |
| BX0214-6 | 1L |
| | 6 x 1L |
| BX0214-7 | 100 ml |
| | 6 x 100 ml |
| | |

Chloroform, DriSolv®

| Trichloromethane | | |
|-------------------------------------|------------|------|
| Stabilized with $\sim 1\%$ | ethanol | |
| CHCI3 | | |
| Specifications: | | |
| Appearance | | |
| Assay (CHCl ₃ + C_2H_5 | | |
| Color (APHA) | | |
| Evaporation residue, p | | |
| Water, ppm | | ≥ 50 |
| ltem | Size | |
| CX1056-6 | 1L | |
| | 6 x 1L | |
| CX1056-7 | 100 ml | |
| | 6 x 100 ml | |
| Assay (GC), % | \geq 99 | |
| Color (APHA) | | ≤ 10 |
| Evaporation residue, p | pm | ≤ 5 |
| Water, ppm | | ≤ 50 |
| | | |
| ltem | Size | |
| CX1057-6 HDS | 1L | |
| | 6 x 1L | |
| CX1057-7 HDS | 100 ml | |
| | 6 x 100 ml | |
| | | |



Not Your Ordinary Bottle Cap!

Chloroform, DriSolv®

Trichloromethane Stabilized with ~ 200ppm alkene CHCI3 Specifications: Appearance...... clear, free from particulates Assay (GC), %.....≥ 99 Color (APHA)..... ≤ 10

| Evaporation residu | e, ppm | ≤ 3 |
|--------------------|--------|------|
| Water, ppm | | ≤ 50 |
| ltem | Size | |
| CX1057-6 | 1L | |
| | 6 x 1L | |
| CX1057-7 | 100 ml | |
| | 6 x 1L | |

| Cyc | lohexane, | DriSolv® |
|-----|-----------|----------|
| | Ц | |

| clear, free f | rom particulates |
|---------------|----------------------|
| | ≥ 99.5 |
| | ≤ 10 |
| e, ppm | ≤ 5 |
| | ≤ 50 |
| | |
| Size | |
| 1L | |
| 6 x 1L | |
| | e, ppm Size 1L |

1,2-Dichlorobenzene, DriSolv® CcH4Cla

| <i>с</i> р. | .40 | 2 | |
|-------------|-----|---|--|
| C | | | |

| Specifications: | | |
|-----------------------------------|---------------|-------------------|
| Appearance | clear, free f | from particulates |
| Assay (GC), % | | ≥ 99 |
| Color (APHA) | | ≤ 10 |
| Evaporation residue, ppm ≤ 3 | | ≤ 3 |
| Water, ppm | | ≤ 50 |
| | | |
| ltem | Size | |
| DX0682-6 | 1L | |
| | 6 x 1L | |

1,2-Dichloroethane, DriSolv®

Ethylene dichloride CH2CICH2CI Specifications: Appearance..... clear, free from particulates Color (APHA) ≤ 10 Evaporation residue, ppm ≤ 3 Water, ppm.... Size ltem DX0794-6 1L 6 x 1L DX0794-7 100 ml 6 x 100 ml

Dichloromethane, DriSolv®

Methylene chloride CH₂Cl₂ Stabilized with an alkene

Specifications:

 ≤ 3

| opeenieacionoi | | |
|----------------|-------------------------------|--|
| Appearance | clear, free from particulates | |
| Assay (GC), % | ≥ 99.8 | |
| Color (APHA) | ≤ 10 | |
| | ie, ppm ≤ 1 | |
| | ≤ 50 | |
| | | |
| ltem | Size | |
| DX0834-6 | 1L | |
| | 6 x 1L | |
| DX0834-7 | 100 ml | |
| | C 100 | |

1,2-Dimethoxyethane, DriSolv®

Ethylene glycol dimethyl ether, Dimethyl glycol CH₃OCH₂CH₂OCH₃ Specifications: Appearance..... clear, free from particulates

| Assav (GC). % | ≥ 99.5 |
|-----------------------------------|--------|
| | ≤ 10 |
| Evaporation residue, ppm ≤ 5 | |
| Water, ppm | ≤ 50 |
| | |
| ltem | Size |
| DX1531-6 | 1L |

6 x 1L

N,N-Dimethylacetamide, DriSolv®

Acetic acid dimethylamide

CH₃CON(CH₃)₂

Specifications:

N.N-Dimethylformamide, DriSoly®

| ingit Dimetingitori | annaci bribori | |
|-------------------------------------|------------------|--------------|
| Formic acid dimet | hylamide | |
| HCON(CH ₃) ₂ | | |
| Specifications: | | |
| Appearance | clear, free from | particulates |
| Assay (GC), % | | ≥ 99.8 |
| Color (APHA) | | ≤ 10 |
| Evaporation residu | ue, ppm | ≤ 5 |
| Water, ppm | | ≤ 50 |
| ltem | Size | |
| DX1727-6 | 1L | |
| | 6 x 1L | |
| DX1727-7 | 100 ml | |
| | 6 x 100 ml | |
| | | |

p-

| -Dioxane, DriSolv® | |
|--|-------------------------------|
| C ₄ H ₈ O ₂ | |
| Unstabilized | |
| Specifications: | |
| Appearance | clear, free from particulates |
| Assay (GC), % | ≥ 99.9 |
| Color (APHA) | ≤ 10 |
| Peroxide, ppm | ≤ 2 (at time of filling) |
| Evaporation residue, ppr | m <u>≤</u> 5 |
| Water, ppm | ≤ 50 |
| lt | Ci |
| Item | Size |
| DX2092-6 | 1L |
| | 6 x 1L |
| | |
| DX2092-7 | 100 ml |

Ethyl Acetate, DriSolv®

| СН ₃ СООС ₂ Н ₅ | | |
|--|-------------------------------|--|
| Specifications: | | |
| Appearance | clear, free from particulates | |
| Assay (GC), % | ≥ 99.9 | |
| Color (APHA) | ≤ 10 | |
| Evaporation residu | e, ppm ≤1 | |
| Water, ppm | ≤ 50 | |
| | | |
| ltem | Size | |
| EX0237-6 | 1L | |
| | 6 x 1L | |
| EX0237-7 | 100 ml | |
| | 6 x 100ml | |

Ethyl Ether, DriSolv®

| (CH ₃ CH ₂) ₂ 0 | | | | |
|---|--------|------|-----|--|
| Stabilized with | \sim | 1ppm | BHT | |

Specifications:

Isopropyl Alcohol, DriSolv®

сн₃снонсн₃

Specifications:

| Appearance | clear, free from particulates |
|----------------------|-------------------------------|
| Assay (GC), % | |
| Color (APHA) | <u><</u> 10 |
| Evaporation residue, | ppm ≤1 |
| Water, ppm \leq 50 | |
| | |
| ltem | Size |
| PX1827-6 | 1L |
| | 6 x 1L |
| PX1827-7 | 100 ml |
| | 6 x 1L |

Methanol, DriSolv® Methyl alcohol СН₃ОН

Specifications:

| Specifications: | |
|-----------------------------------|-------------------------------|
| Appearance | clear, free from particulates |
| Assay (GC), % | ≥ 99.8 |
| Color (APHA) | ≤ 10 |
| Evaporation residue, ppm ≤ 1 | |
| Water, ppm ≤ 50 | |
| | |
| ltem | Size |
| MX0472-6 | 1L |
| | 6 x 1L |
| MX0472-7 | 100 ml |
| | 6 x 100 ml |
| | |

n-Hexane, 95%, DriSolv®

| С ₆ Н ₁₄ | | |
|--------------------------------|-----------------------------|-----|
| Specifications: | | |
| Appearance | clear, free from particulat | tes |
| Assay (n-Hexane) | (GC), %≥ | 95 |
| Color (APHA) | ≤ | 10 |
| Evaporation residu | ie, ppm ≤ | ≤ 3 |
| Water, ppm | ≤ | 50 |
| | | |
| ltem | Size | |
| HX0304-6 | 1L | |
| | 6 x 1L | |
| HX0304-7 | 100 ml | |
| | 6 x 100 ml | |
| | | |

Methyl Sulfoxide, DriSolv®

| Dimethyl sulfoxid | le |
|------------------------------------|-------------------------------|
| (CH ₃) ₂ SO | |
| Specifications: | |
| Appearance | clear, free from particulates |
| Assay (GC), % | |
| Color (APHA) | |
| Evaporation residue | , ppm ≤ 5 |
| Water, ppm | |
| ltem | Size |
| MX1457-6 | 1L |
| | 6 x 1L |
| MX1457-7 | 100 ml |
| | 6 x 100 ml |
| | |

Pentane, DriSolv®

| C5 ^H 12 | | |
|----------------------|--------------------------|-----------|
| Specifications: | | |
| Appearance | clear, free from partici | ulates |
| Assay (GC), % | | ≥ 99 |
| Color (APHA) | | ≤ 10 |
| Evaporation residue, | ppm | ≤ 3 |
| Water, ppm | | ≤ 50 |
| ltem | Size | |
| PX0171-6 | 1L | |
| | 6 x 1L | |
| PX0171-7 | 100 ml | |
| | 6 x 100 ml | |
| | | |

Pyridine, DriSolv[®]

| C ₅ H ₅ N | |
|---------------------------------|-------------------------------|
| Specifications: | |
| Appearance | clear, free from particulates |
| Assay (GC), % | ≥ 99.8 |
| Color (APHA) | ≤ 10 |
| Evaporation residue, | ppm≤ 5 |
| Water, ppm | ≤ 50 |
| | |
| ltem | Size |
| PX2012-6 HDS | 1L |
| | 6 x 1L |
| PX2012-7 HDS | 100 ml |
| | 6 x 100 ml |
| | |

Tetrahydrofuran, DriSolv°

Unstabilized

C₄H₈0

Specifications:

| Appearance | . clear, free from particulates |
|---------------|---------------------------------|
| Assay (GC), % | ≥ 99.9 |
| Color (APHA) | ≤ 10 |
| UV cutoff, nm | |
| Water, ppm | ≤ 50 |

| ltem | Size |
|--------------|------------|
| TX0277-6 HDS | 1L |
| | 6 x 1L |
| TX0277-7 HDS | 100 ml |
| | 6 x 100 ml |

| drofuran, | |
|-----------|--|
| | |

Stabilized with $\sim 25~\text{ppm BHT}$ $C_4 H_8 O$

Specifications:

| Appearance | clear, free from particulates |
|---------------|-------------------------------|
| Assay (GC), % | ≥ 99.9 |
| Color (APHA) | ≤ 10 |
| Water, ppm | ≤ 50 |
| | |
| | Cine |

 Item
 Size

 TX0284-6 HDS
 1L

 6 x 1L
 6 x 1L

 TX0284-7 HDS
 100 ml

 6 x 100 ml
 6 x 100 ml

Toluene, DriSolv®

Methylbenzene

$C_6H_5CH_3$

Specifications:

| Appearance cl | ear, free from particulates |
|--------------------------|-----------------------------|
| Assay (GC), % | ≥ 99.8 |
| Color (APHA) | ≤ 10 |
| Evaporation residue, ppm | ≤ 5 |
| Water, ppm | ≤ 50 |

| Item | Size |
|--------------|------------|
| TX0732-6 HDS | 1L |
| | 6 x 1L |
| TX0732-7 HDS | 100 ml |
| | 6 x 100 ml |

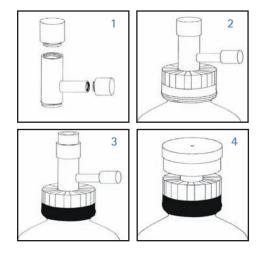
DriSolv® Solvent Bottle Accessories

1. Elbow adapter for use with nitrogen purge. Supplied with plugs and secondary septa to reseal after use. Available as item #692004-2. Replacement plug for bottle top available as item #692012-2.

2. Elbow adapter threaded into bottle cap.

3. Elbow adapter shown with secondary septum, six supplied with item #692004-2. Secondary replacement septa available as item #692004S-2.

4. Drying tube to maintain low moisture level inside the bottle when removing solvent without inert gas flow. Available as item #692010-2.



ELBOW ADAPTER with SEPTUM CAP

For solvent withdrawal under nitrogen purge. Secondary septum cap reduces leakage after primary septum has been punctured.

ltem Size 692004-2 1 unit

SECONDARY REPLACEMENT SEPTA

Pack of ten replacement septa, for use with elbow adapter.

ltem Size 692004S-2 1 pkg

DRYING TUBE

Supplied with 250 g of t.h.e.* desiccant, 100% indicating silica gel item #DX0017 to maintain low moisture content when withdrawing solvent. Other Drying Agents can be used as desired.

ltem 692010–2 Size 1 unit

LARGE PLUG

Replacement plug for bottle top.

| ltem | Size |
|----------|--------|
| 692012-2 | 1 unit |

Extrelut® QE Columns

Extrelut QE Columns are pre-packed with diatomaceous earth for rapid sample cleanup. They are a substitute for liquid-liquid extraction. After pH adjustment (traditional liquid-liquid extraction techniques) and adding sodium chloride (for salting out), the aqueous sample is adsorbed onto the diatomaceous earth in the Extrelut column. Elution of the desired components is then accomplished by elution with a pure or mixture of solvents (e.g. ethyl acetate, diethyl ether, chloroform, toluene, etc.) that would be immiscible with the water adsorbed on its matrix. Using a column version of liquid-liquid extraction is a much easier, quicker way to remove contaminants and allow cleaner, more reproducible LC, GC, and TLC analyses. For papers using these in various protocols, search PubMed on the internet, then search Extrelut on the PubMed site.

It is important to order the correct size column to hold the volume you desire to place on the column. If you have a 10 ml aqueous sample, you need to order a 10 ml Extrelut column. Unlike solid phase extraction (SPE) columns, these columns have no silanols or bonded phase to absorb specific compounds.

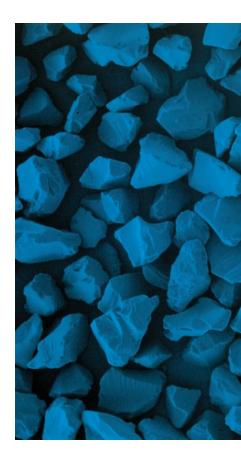
| Catalog No. | Description |
|-------------|--|
| | |
| 901000-1 | Extrelut® QE Column, 0.3 ml. For Sample Preparation |
| 901001-1 | Extrelut® QE Column, 1 ml. For Sample Preparation |
| 901003-1 | Extrelut® QE Column, 3 ml. For Sample Preparation |
| 901005-1 | Extrelut® QE Column, 5 ml. For Sample Preparation |
| 901010-1 | Extrelut® QE Column, 10 ml. For Sample Preparation |
| 901020-1 | Extrelut® QE Column, 20 ml. For Sample Preparation |
| 902050-1 | Extrelut® QE Column, 50 ml. For Sample Preparation |
| 903020-1 | Extrelut® QE Column, Unbuffered, 20 ml. For Sample Preparation |
| 903110-1 | Extrelut® QE Column, Buffered to pH 9.0, 10 ml. For Sample Preparation |
| 903120-1 | Extrelut® QE Column, Buffered to pH 9.0, 20 ml. For Sample Preparation |
| 903220-1 | Extrelut® QE Column, Buffered to pH 4.5, 20 ml. For Sample Preparation |

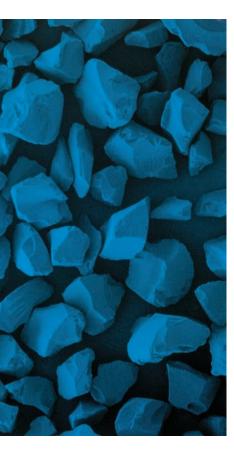
We Begin by Producing the Highest Quality Chromatography Grade Silica ...

Every batch is closely monitored and controlled to provide the pore size, density, pore volume, surface area, and pore size distribution required for chromatographic applications. From these large parent batches, we derive unique silica preparations specific to a variety of classes of Silica Gel 60 products.

LiChrosorb[®]: This analytical grade of irregular materials comes in 5 µm and 10 µm particle sizes for use in HPLC and is available in a wide range of bonded phases. Standardized Silica Gels: These silica gel products are the work horses for Flash chromatography, industrial scale LC and batch adsorption applications. They offer small particle size distribution ranges and are available in three different size ranges: 40-63 µm, 63-200 µm and 200-500 µm.

Geduran[®]: This class of silica gel is made from wider cuts of the same high performance, high quality base material. The wider particle size distribution allows for the same quality at a more competitive cost. It is available in two nominal size ranges: 40-63 µm and 63-200 µm. This class is often selected for less critical Flash chromatography and classical LC applications.





All these products exhibit the same selectivity characteristics as determined by the parent silica, which is also used to manufacture our renowned range of TLC plates. Using EMD Chemicals' TLC plates and EMD Chemicals' silica gels guarantees no selectivity surprises when scaling from TLC to column chromatography.

To extend the range of our irregular silica materials and address some specific needs, we also offer Silica Gel 40 and Silica Gel 100 products which have different mean pore sizes of 40 Angstroms and 100 Angstroms, respectively. The different pore size results in a change in surface areas, pore size distributions, and in the bulk density of the materials which makes them more suited to certain specific applications. However, they are manufactured and tested to the same standards of quality and purity as our Silica Gel 60 products. To complement this range of silicas, we also produce LiChrospher® spherical material for High Performance Preparative Chromatography applications. Because the material is made in very high production volumes, the resulting lot sizes make LiChrospher[®] silica ideally suited for large scale use in the industrial purifications of high quality chemicals and pharmaceuticals. LiChrospher® is available in 60, 100 and 300 Angstrom pore sizes and bonded C8 and C18 silica gel. Our RPselect B is a base deactivated C8 bonded LiChrospher® material for the separation of the basic drug compounds found in the Pharmaceutical industry today. The 300 Angstrom material is ideally suited to small protein and peptide purification.

| Catalog No. | Description | Use | pH* | Size | | |
|-----------------|---|-----|-----|-------|--|--|
| Particle Size 5 | Particle Size 5-40 μm | | | | | |
| 7731-1 | Silica Gel 60 G | TLC | 7.0 | 500g | | |
| 7731-3 | Silica Gel 60 G | TLC | 7.0 | 1kg | | |
| 11677-3 | Silica Gel 60 G** | TLC | 7.0 | 1kg | | |
| 7730-5 | Silica Gel 60 G F ₂₅₄ | TLC | 7.0 | 5kg | | |
| 7741-3 | Silica Gel 60 H F ₂₅₄ + ₃₆₆ | TLC | 7.0 | 1kg | | |
| 7744-1 | Silica Gel 60 HR Extra Pure | TLC | 7.0 | 500g | | |
| 7749-3 | Silica Gel 60 P F_{254} with Gypsum | PLC | 7.0 | 1kg | | |
| 7749-2 | Silica Gel 60 P F ₂₅₄ with Gypsum | PLC | 7.0 | 2.5kg | | |
| | Particle Size <63 μm | | | | | |
| 7729-3 | Silica Gel 60 | TLC | 7.0 | 1kg | | |
| 7729-5 | Silica Gel 60 | TLC | 7.0 | 5kg | | |
| Aluminum Ox | Aluminum Oxide Particle Size 5-40 μm | | | | | |
| 1090-9 | Aluminum Oxide 60 G Neutral | TLC | 7.5 | 25kg | | |
| 1092-1 | Aluminum Oxide 60 G F ₂₅₄ Neutral | TLC | 7.5 | 500g | | |
| 1097-3 | Aluminum Oxide 90, Basic | TLC | 7.5 | 1kg | | |
| 7719-1 | Silica Gel 60 | TLC | 7.0 | 250g | | |
| 7719-3 | Silica Gel 60 | TLC | 7.0 | 1kg | | |
| Additional TLC | Adsorbent Particle Size 50-40 μm | | | | | |
| 8129-1 | Kieselguhr G | TLC | 7.0 | 500g | | |

Adsorbents for Analytical and Preparative Thin Layer Chromatography

Adsorbents for Preparative Column Chromatography (Fe <0.02%, Cl <0.02%)

Mesh Size pH*

Pkg. of

Catalog No. Description

Particle Size 15-40 µm

| 15111-1 | Silica Gel 60 | 350-600 | 7.0 <u>+</u> 0.5 | 500g | |
|--------------------------|--------------------------|---------|------------------|-------|--|
| 15111-3 | Silica Gel 60 | 350-600 | 7.0 <u>+</u> 0.5 | 1kg | |
| | | | | | |
| Particle Size | e 40-63 μm | | | | |
| 9385-3 | Silica Gel 60 | 230-400 | 7.0 <u>+</u> 0.5 | 1kg | |
| 9385-4 | Silica Gel 60 | 230-400 | 7.0±0.5 | 2.5kg | |
| 9385-5 | Silica Gel 60 | 230-400 | 7.0 <u>+</u> 0.5 | 5kg | |
| 9385-9 | Silica Gel 60 | 230-400 | 7.0±0.5 | 25kg | |
| | | | | | |
| Particle Size | e 60-160 μm | | | | |
| 10601-1 | Kieselguhr | | | 500g | |
| | | | | | |
| Particle Size | e 63-200 μm | | | | |
| 10180-3 | Silica Gel 40 | 70-230 | 5.5 <u>+</u> 0.5 | 1kg | |
| 7734-3 | Silica Gel 60 | 70-230 | 7.0 <u>±</u> 0.5 | 1kg | |
| 7734-4 | Silica Gel 60 | 70-230 | 7.0 <u>+</u> 0.5 | 2.5kg | |
| 7734-7 | Silica Gel 60 | 70-230 | 7.0 <u>+</u> 0.5 | 25kg | |
| 7754-1 | Silica Gel 60 Extra Pure | 70-230 | 6.5±0.5 | 500g | |
| 7754-3 | Silica Gel 60 Extra Pure | 70-230 | 6.5 <u>+</u> 0.5 | 1kg | |
| 10184-5 | Silica Gel 100 | 70-230 | 6.5 | 5kg | |
| | | | | | |
| Particle Size 200-500 μm | | | | | |
| 10181-3 | Silica Gel 40 | 35-70 | 5.5 <u>+</u> 0.5 | 1kg | |
| 7733-1 | Silica Gel 60 | 35-70 | 7.0 <u>±</u> 0.5 | 500g | |
| 7733-3 | Silica Gel 60 | 35-70 | 7.0 <u>±</u> 0.5 | 1kg | |
| 7733-5 | Silica Gel 60 | 35-70 | 7.0 <u>±</u> 0.5 | 5kg | |
| 7733-9 | Silica Gel 60 | 35-70 | 7.0±0.5 | 25kg | |
| 10185-1 | Silica Gel 100 | 35-70 | 6.5 | 500g | |
| | | | | - | |
| | | | | | |

*pH of a 10% aqueous suspension

**Mean particle size is 15 µm

Adsorbents for Analytical and Preparative Thin Layer Chromatography

| Catalog No. D | Description | Particle Siz | e Size | | |
|------------------------|-------------------|--------------|--------|--|--|
| 10167-3 S | ilica Gel 60 RP18 | 40-63 µm | 100g | | |
| | | | | | |
| Geduran® Silica Gel 60 | | | | | |
| Catalog No. D | Description | pН | Size | | |

Particle Size 40-63 μm

| 11567-2 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 500g |
|---------|---------------|------------------|-------|
| 11567-3 | Silica Gel 60 | 7.0±0.5 | 1kg |
| 11567-6 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 2kg |
| 11567-4 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 2.5kg |
| 11567-5 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 5kg |
| 11567-1 | Silica Gel 60 | 7.0 <u>+</u> 0.5 | 25kg |
| | | | |

Particle Size 63-200 µm

| 110832-3 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 1kg |
|----------|---------------|------------------|-------|
| 110832-4 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 2.5kg |
| 110832-1 | Silica Gel 60 | 7.0 <u>±</u> 0.5 | 25kg |
| | | | |

Alumina Adsorbents for Preparative Column Chromatography

| Catalog No. | Description | рН | Size |
|-------------|----------------------------|------------------|------|
| 1067-3 | Aluminum Oxide 60, Basic | 9.0 <u>+</u> 0.5 | 1kg |
| 1067-6 | Aluminum Oxide 60, Basic | 9.0 <u>+</u> 0.5 | 2kg |
| 1076-3 | Aluminum Oxide 90, Basic | 9.0 <u>+</u> 0.5 | 1kg |
| 1076-6 | Aluminum Oxide 90, Basic | 9.0±0.5 | 2kg |
| 1077-2 | Aluminum Oxide 90, Neutral | 7.3 <u>±</u> 0.5 | 1kg |
| 1077-6 | Aluminum Oxide 90, Neutral | 7.3 <u>±</u> 0.5 | 2kg |
| 1077-7 | Aluminum Oxide 90, Neutral | 7.3±0.5 | 20kg |
| 1078-3 | Aluminum Oxide 90, Acidic | 4.0±0.5 | 1kg |
| 1078-6 | Aluminum Oxide 90, Acidic | 4.0 <u>+</u> 0.5 | 2kg |

Spherical Preparative Chromatography - Sorbents

| Catalog No. | Description | Particle Siz | e Size |
|-------------|--|--------------|--------|
| 19654-5 | LiChrospher [®] 60 | 12µm | 100g |
| 19654-7 | LiChrospher [®] 60 | 12µm | 1kg |
| 19655-4 | LiChrospher [®] 60 RPselect B | 12µm | 100g |
| 19655-5 | LiChrospher [®] 60 RPselect B | 12µm | 500g |
| 19656-5 | LiChrospher [®] 100 RP18 | 12µm | 100g |
| 19656-4 | LiChrospher [®] 100 RP18 | 12µm | 500g |
| 19662-1 | LiChrospher [®] 300 RP18 | 12µm | 100g |
| 19662-4 | LiChrospher [®] 300 RP18 | 12µm | 500g |
| 11024-5 | LiChrospher [®] 60 | 15µm | 100g |
| 11024-7 | LiChrospher [®] 60 | 15µm | 1kg |
| 11023-5 | LiChrospher [®] 60 RPselect B | 15µm | 100g |
| 11023-4 | LiChrospher [®] 60 RPselect B | 15µm | 500g |
| 11022-5 | LiChrospher [®] 100 RP18 | 15µm | 100g |
| 11022-4 | LiChrospher [®] 100 RP18 | 15µm | 500g |
| 19659-1 | LiChrospher [®] 300 RP18 | 15µm | 100g |
| 19659-4 | LiChrospher [®] 300 RP18 | 15µm | 500g |



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