

## Product Information

**MEK2, active, GST-tagged, human  
PRECISIO® Kinase  
recombinant, expressed in Sf9 cells**

Catalog Number **M1198**  
Lot Number 021M0608  
Storage Temperature  $-70\text{ }^{\circ}\text{C}$

Synonyms: MAP2K2, MKK2, PRMK2, MAPKK2

### Product Description

MEK2 is a member of the MAPK kinase (MAPKK) family of signaling protein kinases. MEK2 is a dual-specificity kinase that activates the extracellular signal-regulated kinase (ERK) and mitogen-activated protein (MAP) kinase upon agonist binding to receptors. MEK2 plays a key role in the Ras/Raf/MEK/ERK mitogen-activated protein kinase (MAPK) signaling pathways.<sup>1</sup> Approximately 30% of all human cancers have a constitutively activated MAPK pathway and constitutive activation of MEK2 results in cellular transformation. The ERK/MAP kinase cascade regulates cell growth and differentiation.<sup>2</sup>

This recombinant product was expressed by baculovirus in Sf9 insect cells using an N-terminal GST-tag. The gene accession number is NM 030662. It is supplied in 50 mM Tris-HCl, pH 7.5, with 150 mM NaCl, 0.25 mM DTT, 0.1 mM EGTA, 0.1 mM EDTA, 0.1 mM PMSF, and 25% glycerol.

Molecular mass: ~71 kDa

Purity:  $\geq 70\%$  (SDS-PAGE, see Figure 1)

Specific Activity: 190–258 nmole/min/mg (see Figure 2)

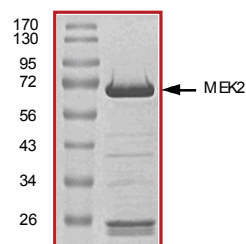
### Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

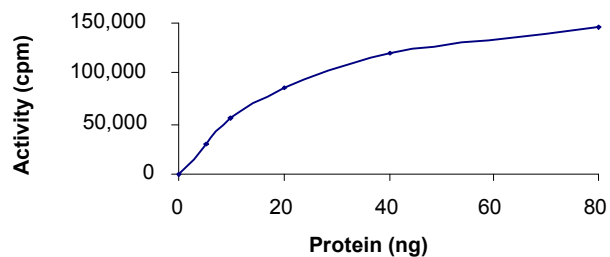
### Storage/Stability

The product ships on dry ice and storage at  $-70\text{ }^{\circ}\text{C}$  is recommended. After opening, aliquot into smaller quantities and store at  $-70\text{ }^{\circ}\text{C}$ . Avoid repeated handling and multiple freeze/thaw cycles.

**Figure 1.**  
SDS-PAGE Gel of Lot Number 021M0608:  
>90% (densitometry)



**Figure 2.**  
Specific Activity of Lot Number 021M0608:  
224 nmole/min/mg



### Procedure

#### Preparation Instructions

Kinase Assay Buffer – 25 mM MOPS, pH 7.2, 12.5 mM glycerol 2-phosphate, 25 mM  $\text{MgCl}_2$ , 5 mM EGTA, and 2 mM EDTA. Just prior to use, add DTT to a final concentration of 0.25 mM.

Kinase Dilution Buffer – Dilute the Kinase Assay Buffer 5-fold with a 50 ng/ $\mu\text{l}$  BSA solution.

Kinase Solution – Dilute the active MEK2 (0.1 µg/µl) with Kinase Dilution Buffer to the desired concentration. **Note:** The lot-specific specific activity plot may be used as a guideline (see Figure 2). It is recommended the researcher perform a serial dilution of active MEK2 kinase for optimal results.

10 mM ATP Stock Solution – Dissolve 55 mg of ATP in 10 ml of Kinase Assay Buffer. Store in 200 µl aliquots at –20 °C.

γ-<sup>32</sup>P-ATP Assay Cocktail (250 µM) – Combine 5.75 ml of Kinase Assay Buffer, 150 µl of 10 mM ATP Stock Solution, 100 µl of γ-<sup>32</sup>P-ATP (1 mCi/100 µl). Store in 1 ml aliquots at –20 °C.

Substrate Solution – Inactive ERK2 (0.2 µg/ml); Myelin Basic Protein (MBP) diluted in water at a final concentration of 1 mg/ml.

1% phosphoric acid solution – Dilute 10 ml of concentrated phosphoric acid to a final volume of 1 L with water.

#### Kinase Assay

This assay involves the use of the <sup>32</sup>P radioisotope. All institutional guidelines regarding the use of radioisotopes should be followed.

1. Thaw the active MEK2, Kinase Assay Buffer, Inactive ERK2, Substrate Solution, and Kinase Dilution Buffer on ice. The γ-<sup>32</sup>P-ATP Assay Cocktail may be thawed at room temperature.
2. In a pre-cooled microcentrifuge tube, prepare an activation mixture with a volume of 20 µl:
  - 5 µl of Kinase Solution
  - 10 µl of Inactive ERK2 (0.2 µg/µl)
  - 5 µl of Kinase Assay Buffer
3. Start the activation reaction by adding 5 µl of 250 µM ATP and incubate in a water bath at 30 °C for 15 minutes.
4. In a microcentrifuge tube, add the following solutions to a volume of 20 µl:
  - 5 µl of activated mixture (step 3)
  - 5 µl of MBP Substrate Solution
  - 10 µl of cold water (4 °C)
5. Set up a blank control as outlined in step 4, substituting 5 µl of cold water (4 °C) for the Substrate Solution.
6. Initiate each reaction with the addition of 5 µl of the γ-<sup>32</sup>P-ATP Assay Cocktail, bringing the final reaction volume to 25 µl. Incubate the mixture in a water bath at 30 °C for 15 minutes.

7. After the 15 minute incubation, stop the reaction by spotting 20 µl of the reaction mixture onto an individually pre-cut strip of phosphocellulose P81 paper.
8. Air dry the pre-cut P81 strip and sequentially wash in the 1% phosphoric acid solution with constant gentle stirring. It is recommended the strips be washed a total of 3 times of ~10 minutes each.
9. Set up a radioactive control to measure the total γ-<sup>32</sup>P-ATP counts introduced into the reaction. Spot 5 µl of the γ-<sup>32</sup>P-ATP Assay Cocktail on a pre-cut P81 strip. Dry the sample for 2 minutes and read the counts. Do not wash this sample.
10. Count the radioactivity on the P81 paper in the presence of scintillation fluid in a scintillation counter.
11. Determine the corrected cpm by subtracting the blank control value (see step 3) from each sample and calculate the kinase specific activity

#### Calculations:

1. Specific Radioactivity (SR) of ATP (cpm/nmole)

$$SR = \frac{\text{cpm of 5 } \mu\text{l of } \gamma\text{-}^{32}\text{P-ATP Assay Cocktail}}{\text{nmole of ATP}}$$

$$\begin{aligned} &\text{cpm} - \text{value from control (step 7)} \\ &\text{nmole} - 1.25 \text{ nmole (5 } \mu\text{l of 250 } \mu\text{M ATP} \\ &\text{Assay Cocktail)} \end{aligned}$$

2. Specific Kinase Activity (SA) (nmole/min/mg)

$$\text{nmole/min/mg} = \frac{\Delta\text{cpm} \times (25/20)}{SR \times E \times T}$$

$$\begin{aligned} SR &= \text{specific radioactivity of the ATP (cpm/nmole ATP)} \\ \Delta\text{cpm} &= \text{cpm of the sample} - \text{cpm of the blank (step 3)} \\ 25 &= \text{total reaction volume} \\ 20 &= \text{spot volume} \\ T &= \text{reaction time (minutes)} \\ E &= \text{amount of enzyme (mg)} \end{aligned}$$

#### **References**

1. Shuichan, X. et al., Mol. Endocrinol., **11**, 1618-1625 (1997).
2. Louis-François, B, et al., Mol. Cellular Biol., **23**, 4778-4787 (2003).

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