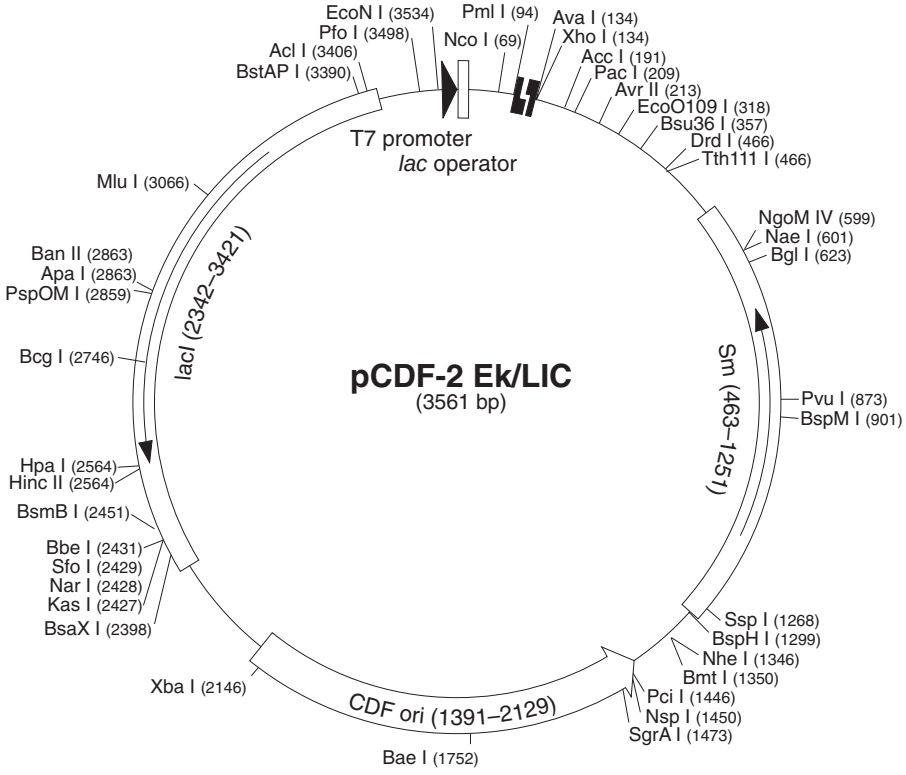
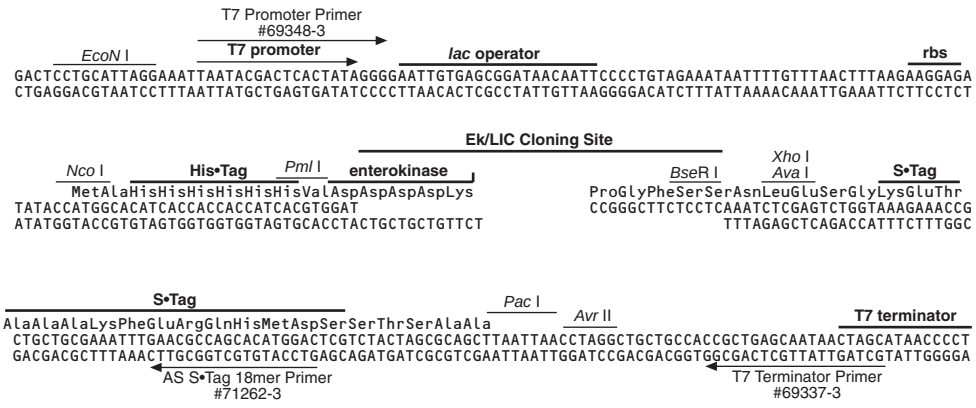
pCDF-2 Ek/LIC Vector

	Cat. No.
pCDF-2 Ek/LIC Cloning Kit	71337-3

pCDF-2 Ek/LIC sequence landmarks	
T7 promoter	3545–3561
T7 transcription start	1
His•Tag® coding sequence	77–94
Ek/LIC cloning site	101–129
S•Tag™ coding sequence	146–190
T7 terminator	242–289
aadA (SmR) coding sequence	463–1251
CDF origin	1391–2129
lacI coding sequence	2342–3421

The pCDF-2 Ek/LIC vector is prepared for rapid, directional cloning of PCR-amplified DNA for high-level expression of polypeptides. The vector carries a T7*lac* promoter to control transcription, a replication origin derived from CloDF13, and streptomycin/spectinomycin antibiotic resistance. Using specifically designed primers for amplification and the pCDF-2 Ek/LIC Cloning Kit (71337-3), inserts can be efficiently cloned without the need for restriction digestion or ligation. Fusion proteins contain an N-terminal cleavable His•Tag® sequence. Fusion to an optional C-terminal S•Tag™sequence can also be created for detection, purification and quantification of fusion proteins. pCDF-2 is compatible with pET vectors (ColE1 origin), with pRSF vectors (RSF1030 replication origin), and pACYC plasmids (P15A replication origin) carrying compatible antibiotic resistance markers. Unique sites are shown on the circle map. Sequencing can be performed using the T7 Promoter Primer (Cat. No. 69348-3)and AS S•Tag 18 mer Primer (Cat. No. 71262-3) or T7 Terminator Primer (Cat. No. 69337-3).





pCDF-2 Ek/LIC cloning/expression region

pCDF-2 Ek/LIC Restriction Sites

TB394 0903

Enzyme	# Sites	Locations				
AccI	1	191				
AclI	1	3406				
AflIII	2	1446	3066			
AgeI	1	346				
ApaI	1	2863				
ApaLI	2	973	3086			
AseI	3	2324	2383	3544		
AvaI	1	134				
AvrII	1	213				
BaeI	1	1752				
BanI	3	2297	2427	3146		
BanII	1	2863				
BbeI	1	2431				
BbsI	2	2581	2920			
BceAI	6	1132	1176	1537	1648	2582
		3209				
BcgI	1	2746				
BciVI	3	1301	1667	2614		
BclI	2	1050	3052			
BglI	1	623				
BlpI	2	231	940			
Bme1580I	3	977	2863	3090		
BmrI	7	604	613	709	1898	2268
		2908	3145			
BmtI	1	1350				
BpmI	3	1002	2745	3234		
BpuEI	4	295	1854	2095	2258	
BsaAI	2	94	495			
BsaHI	3	1196	2428	3111		
BsaWI	8	331	346	782	1534	1670
		1818	2244	2747		
BsaXI	1	2398				
BseRI	1	115				
BseYI	3	1769	2532	2667		
BsgI	2	3021	3221			
BsiEI	4	405	873	1360	2287	
BsiHKAI	3	977	1989	3090		
BsmAI	6	1301	1372	2451	2838	2964
		3369				
BsmBI	1	2451				
BsmFI	2	692	1453			
Bsp1286I	4	977	1989	2863	3090	
BspCNI	3	223	310	2487		
BspHI	1	1299				
BspMI	1	901				
BsrBI	3	13	1305	2144		
BsrDI	4	612	905	2659	3025	
BsrFI	5	346	599	652	1473	3380
BssHII	2	524	2655			
BstAPI	1	3390				
BstEII	2	1087	2884			
BstXI	3	3020	3143	3272		
BstYI	7	421	767	1607	2106	2117
		2290	3502			
Bsu36I	1	297				
BtgI	4	69	955	1425	2006	
BtsI	4	323	1137	2339	2707	
DraIII	2	498	1118			
DrdI	1	406				
EaeI	3	416	1161	2392		
EarI	3	1259	2174	3449		
EciI	4	790	1486	1673	3281	
Eco57I	2	1108	2206			
Eco57MI	5	1002	1108	2206	2745	3234
EcoNI	1	3534				
EcoO109I	1	258				
HaeII	8	610	759	1189	1394	1714
		2431	2674	3455		
HincII	1	2564				
HpaI	1	2564				
KasI	1	2427				
MluI	1	3066				

Enzyme	# Sites	Locations				
MslI	5	96	980	2700	2730	3018
NaeI	1	601				
NarI	1	2428				
NcoI	1	69				
NgoMIV	1	599				
NheI	1	1346				
NspI	1	1450				
PacI	1	209				
PciI	1	1446				
PflMI	3	94	181	3491		
PfoI	1	3498				
PmlI	1	94				
PshAI	1	111				
PspOMI	1	2859				
PvuI	1	873				
PvuII	2	2377	2470			
SfcI	3	29	993	3557		
SfoI	1	2429				
SgrAI	1	1473				
SmlI	5	134	274	1833	2110	2273
SspI	1	1268				
StyI	5	69	213	253	474	816
TaqII	2	1346	2273			
TspGWI	2	972	1414			
Tth111I	1	406				
XbaI	1	2146				
XcmI	4	1423	2681	2699	3215	
XhoI	1	134				

Enzymes that do not cut pCDF-2 Ek/LIC:

AarI	AatII	Acc65I	AfeI	AflII
AhdI	AleI	AloI	AlwNI	AscI
AsiSI	BamHI	BbvCI	BfrBI	BglII
BmgBI	BpiI	Bpu10I	BsaBI	BsaI
BsiWI	BsmI	BspEI	BsrGI	BssSI
BstBI	BstZ17I	ClaI	DraI	EagI
EcoICRI	EcoRI	EcoRV	FalI	FseI
FspAI	FspI	HindIII	KpnI	MfeI
MscI	NdeI	NotI	NruI	NsiI
PmeI	PpiI	PpuMI	PsiI	PsrI
PstI	RsrII	SacI	SacII	SalI
SanDI	SapI	SbfI	ScaI	SexAI
SfiI	SmaI	SnaBI	SpeI	SphI
SrfI	StuI	SwaI	TatI	XmaI
XmnI	ZraI			