

Product Information

CompoZr® Custom Zinc Finger Activators CompoZr Custom ZFA, Rep, Epigenetic Mod

Catalog Numbers CSTZFACT-1KT
CZFA2136-1KT (ZFA targeting human SOX2)
CZFA17184-1KT (ZFA targeting human OCT4)
CZFA1504-1KT (ZFA targeting human KLF4)
CZFA1041-1KT (ZFA targeting human c-MYC)

Technical Bulletin

Important

This protocol is optimized for use with TransIT LT®-1 (Mirus Bio)

Other suggested transfection reagents include:

TransIT®- 2020 (Mirus Bio)

Lipofectamine® 2000 (Invitrogen)

Any transfection reagent may require additional optimization of the protocol

Product Description

Various classes of naturally occurring DNA-binding molecules, including zinc-finger proteins (1,2), triplex forming oligos (TFOs) (3), meganucleases (4), transcription activator-like effectors (TALEs) (5,6), and the recently discovered cas9 proteins (7,8), have been engineered to bind sequence-specific endogenous target sites. Such DNA-specific binding proteins, when coupled to various functional domains, are powerful tools for modifying the genome or regulating gene expression. Zinc finger proteins have been highly characterized for such uses (1,2). These modular proteins consist of multiple zinc-finger domains, each recognizing 3 base pairs of DNA linked together to generate a protein that binds specific DNA sequences. When fused to a transcriptional activation domain (such as VP16 from the herpes simplex virus), these engineered transcription factors (ZF-TFs) effectively upregulate target gene expression when delivered to cells (9-14).

The zinc-finger domain for each ZF-TF consists of either five or six zinc-finger subunits, resulting in a ZF-TF with either a 15 or 18 base-pair DNA recognition site. Each 5 or 6 ZF coding sequence assembly is cloned between an N-terminal nuclear localization signal and a C-terminal NF-κB p65 activation domain to generate a series of ZF-TF expression constructs. The resulting fusion protein is designed to bind to the specific target sequence on either the forward or reverse strand of DNA. The ZF-TFs are then screened in HEK293 cells (for human gene targets) or Neuro 2A cells (for murine targets) by transfection with TransIT-LT1 (Mirus Bio). Measurement of target gene mRNA is used to determine whether designed ZF-TFs successfully upregulate the target gene's expression (Figure 1).

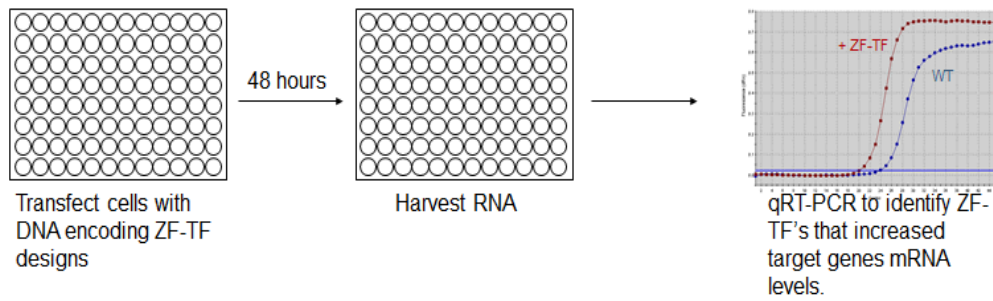


Figure 1. Schematic of the ZF-TF Validation Assay

Cells are harvested 48 hours post-transfection, and total RNA is isolated using RNeasy Plus 96 Kit (Qiagen) or RNeasy Plus Mini Kit (Qiagen). The levels of target gene mRNA and *PPIA* (cyclophilin A) endogenous control (Applied BioSystems) mRNA are measured by qRT-PCR using Quantitative RT-PCR ReadyMix (Sigma-Aldrich Biotechnology), following the manufacturer's protocol. qRT-PCR analysis is performed using the Mx3000P (Stratagene).

Unvalidated ZFAs are also available. For the unvalidated ZFAs, up to 12 ZFA plasmids (with the p65 activation domain) will be shipped. Please note that the qRT-PCR validation assay is not included with the unvalidated ZFA products.

Optimizing ZF-TF activity via effector domain selection

The effector domain linked to a ZF-TF can have a large influence on the level of transcriptional activation, repression, or modification achieved by such artificial transcription factors (16). Using ZF-TFs identified in the screening method above, the end user can further optimize targeted gene activation by fusing different effector domains to the zinc-finger domains in place of the NF- κ B p65 activation subunit.

In order to demonstrate the varied efficacy of each domain in the context of individual target gene activation, ZF-TFs targeting *OCT4* (CZFA17184-1KT), *SOX2*, (CZFA2136-1KT), *KLF4* (CZFA1504-1KT), and *c-MYC* (CZFA1041-1KT) were tested with p65, VP16, VP64 or 2Xp65 (two tandem copies of the p65 domain) activation domains (Figure 2).

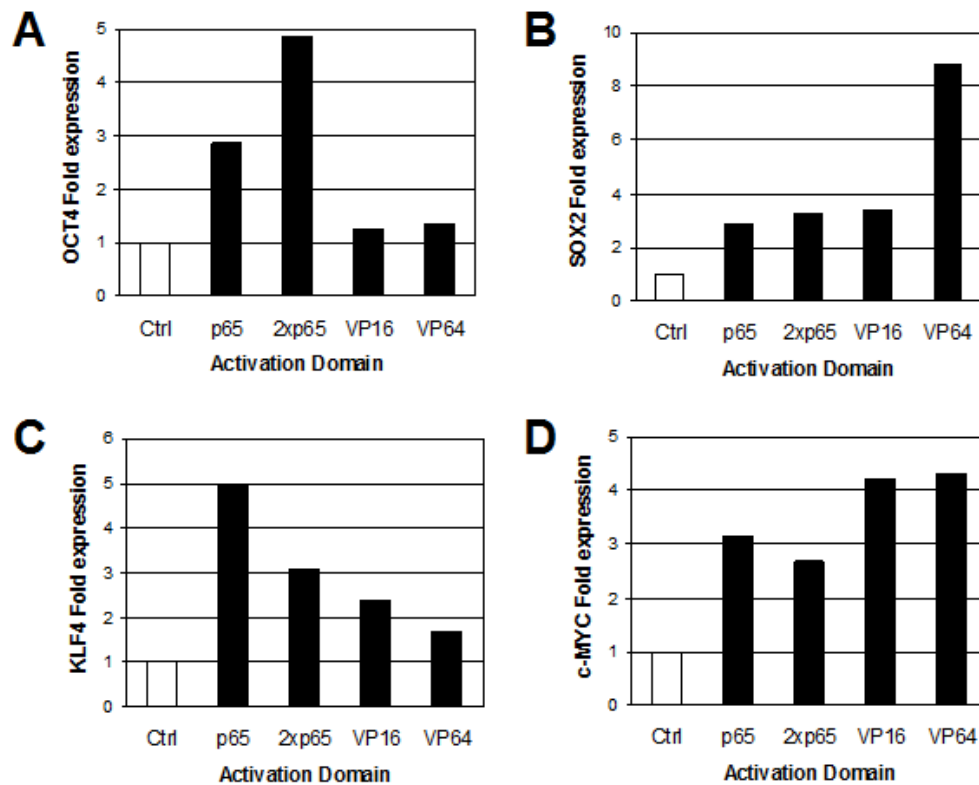


Figure 2. Use of various effector domains with ZFPs to upregulate mRNA expression.

ZF-TF Plasmid Design and Cloning Options

ZF-TF molecules are delivered in plasmid format, in the vector backbone shown below (Figure 3). Expression of ZF-p65 is driven by the CMV promoter, along with an upstream GFP-2A cDNA, allowing for visual selection of cells that contain your plasmid of interest. This in turn allows for more accurate expression data analysis for both in vivo and in vitro experiments.

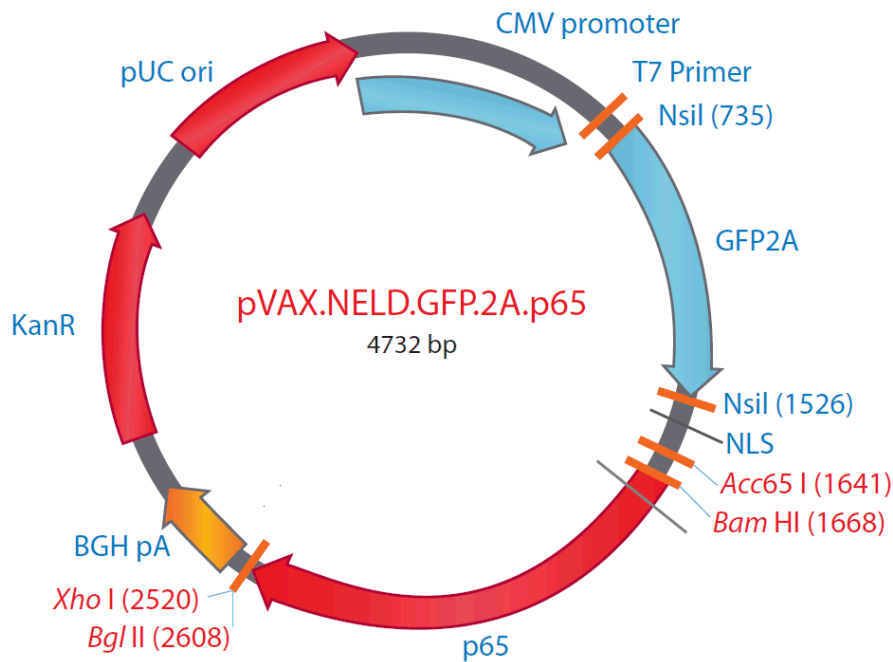


Figure 3. Map of the ZF-TF cloning vector (with p65 effector domain).

Cloning to alternate and remove effector domains

Standard format for ZF-TF delivery is via the plasmid containing the p65 activation domain. Zinc finger motifs are cloned 5' of this sequence using *Acc65I* and *BamHI* sites (see Figure 3 for reference). The necessary plasmid backbones that contain additional activation domains (VP16, VP64, or 2xp65), repressor domain (KRAB), or epigenetic modification domains (p300 or G9A) are provided along with the standard p65 plasmid. To manipulate the domain on each ZF-TF, a simple transfer of the zinc finger module to the destination plasmid will be required, using *Acc65I* and *BamHI* to perform the restriction digestions. Upon ligation into the destination plasmid, the zinc-finger module will now be in frame with the downstream activation domain, resulting in a ZF-TF fusion protein. In addition, a zinc-finger only control (lacking the transactivating domain) can be created by simple digest of the original ZF-TF-p65 plasmid with *BamHI* and *BglII*, followed by purification and intramolecular ligation of the larger DNA fragment containing the zinc-finger sequence. These ends are cohesive and compatible and can be ligated together to form your domain drop out vector, which can be used for control experiments.

General Guidelines for Transfection

Given that the ZF-TF product is delivered in plasmid format, reagents should be immediately ready for small-scale transfection, unless scale up and alternate domain optimization is desired. While this product has been optimized using TransIT LT®-1 (Mirus Bio), use of a variety of transfection methods and reagents is possible. It is expected that the end user will optimize transfection conditions based on cell type of interest and any downstream assay constraints. For best results, Sigma-Aldrich recommends a starting range 0.5 to 1.5 micrograms of ZF-TF plasmid per well in either 24- or 12- well plates, containing 50,000 to 100,000 cells per well. Optimization within this range should generally yield at least a 2-fold change in target gene expression for a validated ZF-TF. It is assumed that these results may vary based on cell type and transfection method of choice.

Precautions and Disclaimer

This product is for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Storage and Stability

Recommended storage of DNA is -20C. Bacterial glycerol stocks should be stored at -80C.

RNases are ubiquitous and very stable proteins, which are a primary concern for any researcher attempting to manipulate RNA. Care must be taken not to introduce RNases. It is recommended to use RNase-free pipette tips, preferably those having an aerosol barrier, to wear latex gloves and change them frequently, and keep bottles and tubes closed when not in use.

Kit components

For each custom order, the zinc finger corresponding to the target of choice will be provided in the following format: ZF + C-terminal NF- κ B p65. Up to three ZFAs will be provided. Additional cloning vectors are provided according to the table below.

Reagent Description	Catalog Number	CSTZFACT – 1KT	CSTZFACT – 1KT Upgrade
pZFA1	D1ZFA	1 EA	1 EA
pZFA2	D2ZFA	None or 1 EA	None or 1 EA
pZFA3	D3ZFA	None or 1 EA	None or 1 EA
pZFA-GFP- ν p16	ZFG0002	1 EA	1 EA
pZFA-GFP- ν p64	ZFG0003	1 EA	1 EA
pZFA-GFP-2xp65	ZFG0004	1 EA	1 EA
pZFE-GFP-p300*	ZFG0005	N/A	1 EA
pZFE-GFP-G9A*	ZFG0006	N/A	1 EA
pZFR-GFP-KRAB*	ZFG0007	N/A	1 EA

*The vectors that contain the p300, G9A and KRAB domains are available as an optional upgrade to the custom ZFA kit.

Note: For the unvalidated ZFAs, up to 12 ZFA plasmids (with the p65 activation domain) will be shipped. Please note that the qRT-PCR validation assay is not included with the unvalidated ZFA products.

Reagents and Equipment Recommended But Not Provided

Nucleofection[®] reagents and instrument or Electroporation reagents and instrument or TransIT[®]-mRNA Transfection Reagent (Mirus Bio Catalog Number MIR 2225, LLC)

Agarose

Hank's Balanced Salt Solution (HBSS, Catalog Number H6648)

DirectLoad 1 kb DNA Ladder (Catalog Number D3937)

Quantitative RT-PCR Ready Mix (Catalog Number QRO200)

References

1. Pavletich, N.P. and Pabo, C.O. (1991) Zinc finger-DNA recognition: crystal structure of a Zif268-DNA complex at 2.1 Å. *Science*, **252**, 809-817.
2. Klug, A. (2010) The discovery of zinc fingers and their applications in gene regulation and genome manipulation. *Annu Rev Biochem*, **79**, 213-231.
3. Duca, M., Vekhoff, P., Oussedik, K., Halby, L. and Arimondo, P.B. (2008) The triple helix: 50 years later, the outcome. *Nucleic Acids Res*, **36**, 5123-5138.
4. Grizot, S., Smith, J., Daboussi, F., Prieto, J., Redondo, P., Merino, N., Villate, M., Thomas, S., Lemaire, L., Montoya, G. *et al.* (2009) Efficient targeting of a SCID gene by an engineered single-chain homing endonuclease. *Nucleic Acids Res*, **37**, 5405-5419.
5. Moscou, M.J. and Bogdanove, A.J. (2009) A simple cipher governs DNA recognition by TAL effectors. *Science*, **326**, 1501.
6. Zhang, F., Cong, L., Lodato, S., Kosuri, S., Church, G.M. and Arlotta, P. (2011) Efficient construction of sequence-specific TAL effectors for modulating mammalian transcription. *Nat Biotechnol*, **29**, 149-153.
7. Cong, L., Ran, F.A., Cox, D., Lin, S., Barretto, R., Habib, N., Hsu, P.D., Wu, X., Jiang, W., Marraffini, L.A. *et al.* (2013) Multiplex Genome Engineering Using CRISPR/Cas Systems. *Science*.
8. Mali, P., Yang, L., Esvelt, K.M., Aach, J., Guell, M., Dicarlo, J.E., Norville, J.E. and Church, G.M. (2013) RNA-Guided Human Genome Engineering via Cas9. *Science*.
9. Rebar, E.J., Huang, Y., Hickey, R., Nath, A.K., Meoli, D., Nath, S., Chen, B., Xu, L., Liang, Y., Jamieson, A.C. *et al.* (2002) Induction of angiogenesis in a mouse model using engineered transcription factors. *Nat Med*, **8**, 1427-1432.
10. Zhang, L., Spratt, S.K., Liu, Q., Johnstone, B., Qi, H., Raschke, E.E., Jamieson, A.C., Rebar, E.J., Wolffe, A.P. and Case, C.C. (2000) Synthetic zinc finger transcription factor action at an endogenous chromosomal site. Activation of the human erythropoietin gene. *J Biol Chem*, **275**, 33850-33860.
11. Beerli, R.R., Segal, D.J., Dreier, B. and Barbas, C.F., 3rd. (1998) Toward controlling gene expression at will: specific regulation of the erbB-2/HER-2 promoter by using polydactyl zinc finger proteins constructed from modular building blocks. *Proc Natl Acad Sci U S A*, **95**, 14628-14633.
12. Yokoi, K., Zhang, H.S., Kachi, S., Balaggan, K.S., Yu, Q., Guschin, D., Kunis, M., Surosky, R., Africa, L.M., Bainbridge, J.W. *et al.* (2007) Gene transfer of an engineered zinc finger protein enhances the anti-angiogenic defense system. *Mol Ther*, **15**, 1917-1923.
13. Liu, P.Q., Rebar, E.J., Zhang, L., Liu, Q., Jamieson, A.C., Liang, Y., Qi, H., Li, P.X., Chen, B., Mendel, M.C. *et al.* (2001) Regulation of an endogenous locus using a panel of designed zinc finger proteins targeted to accessible chromatin regions. Activation of vascular endothelial growth factor A. *J Biol Chem*, **276**, 11323-11334.
14. Bartsevich, V.V., Miller, J.C., Case, C.C. and Pabo, C.O. (2003) Engineered zinc finger proteins for controlling stem cell fate. *Stem Cells*, **21**, 632-637.
15. Bultmann, S., Morbitzer, R., Schmidt, C.S., Thanisch, K., Spada, F., Elsaesser, J., Lahaye, T. and Leonhardt, H. (2012) Targeted transcriptional activation of silent oct4 pluripotency gene by combining designer TALEs and inhibition of epigenetic modifiers. *Nucleic Acids Res*, **40**, 5368-5377.
16. Sera, T. (2009) Zinc-finger-based artificial transcription factors and their applications. *Adv Drug Deliv Rev*, **61**, 513-526.

License Agreement

This Product and its use are the subject of one or more of the following patents controlled by Sangamo BioSciences, Inc.: U.S. Patent Nos. 6,534,261, 6,607,882, 6,746,838, 6,794,136, 6,824,978, 6,866,997, 6,933,113, 6,979,539, 7,013,219, 7,030,215, 7,220,719, 7,241,573, 7,241,574, 7,585,849, 7,595,376, 6,903,185, 6,479,626, US20030232410, US20090203140 and corresponding foreign patent applications and patents.

BEFORE OPENING OR USING THIS PRODUCT, PLEASE READ THE TERMS AND CONDITIONS SET FORTH IN THIS LICENSE AGREEMENT. YOUR USE OF THIS PRODUCT SHALL CONSTITUTE ACKNOWLEDGMENT AND ACCEPTANCE OF THESE TERMS AND CONDITIONS. If you do not agree to use this Product pursuant to the terms and conditions set out in this License Agreement, please contact Sigma Technical Services within ten days of receipt to return the unused and unopened Product for a full refund; provided, however, that custom-made Products may not be returned for a refund.

The purchase of this Product conveys to you, the buyer, the non-transferable right to use the purchased Product for Licensed Research Use (see definition below) subject to the conditions set out in this License Agreement. If you wish to use this Product for any purpose other than Licensed Research Use, you must first obtain an appropriate license (see information set out below).

This Product may not be used for any purpose other than Licensed Research Use. Your right to use this Product for Licensed Research Use is subject to the following conditions and restrictions:

1. "Licensed Research Use" means any use for research purposes, other than:
 - (a) Licensing, selling, distributing, or otherwise providing Modified Animals to any third party other than Sigma and its affiliates as provided herein: provided however, that you may provide Modified Animals to researchers within your research organization located at the same research facility or campus. A "Modified Animal" means an animal having a genomic modification at the target site that results from Customer's use of the Product. Modified Animal includes but is not limited to (a) heterozygotes and mosaic animals, (b) the descendants of Modified Animals, (c) animals created from the breeding of Modified Animals with other animals, and (d) animals created by the Customer which contain and/or incorporate genetic information derived from Modified Animals. You may not breed or otherwise develop Modified Animals in a quantity in excess of the following without a further license from Sigma:
 - Danio, Drosophila, or Xenopus: unlimited animals
 - Mice, Rats, or Rabbits: 500 animals
 - Domesticated farm animals: 10 animals
 For questions about animals not listed above, please contact Sigma.
 - (b) GMP production of therapeutic, diagnostic, prophylactic or other medicinal Products intended for use in humans or non-human animals, or any other industrial use solely to the extent involving commercial sale of a Product or service. If a molecule or any derivative of such molecule is used in or administered to humans, then the production of such molecule shall be deemed to be GMP production and therefore in violation of this License Agreement;
 - (c) the use of the Product or a direct derivative (a modified cell or Modified Animal resulting from use of the Product) in the screening or testing of more than 10,000 distinct compounds (high throughput screening);
 - (d) use for gene targeting and/or gene regulation to modify the genome of a plant cell, plant, or plant cell culture (in each case, whether constituting or derived from a vascular or non-vascular plant), or alter the nucleic acid or protein expression in a plant cell, plant, or plant cell culture. "Non-vascular" plants shall include but not be limited to algae, moss, and fungi; and
 - (e) modification or reverse-engineering of the Product in any way or creating any derivatives or sequence variants thereof.

2. You may not transfer the Product, its components, or any materials made through the use of this Product, including Modified Animals, to any third party without prior written approval of Sigma and without the transferee entering into a use license with Sigma. Notwithstanding the foregoing, the Product or materials made through use of the Product may be transferred by you to your legal affiliates or bonafide third party contractors performing paid work on your behalf, with the exception of creation of Modified Animals, provided the use by such third party contractors is limited to performance of work for you.
3. You may not transfer the Product or materials made through use of the Product to third party contractors performing paid work on your behalf for the purposes of creation of Modified Animals.
4. Your right to use the Product will terminate immediately if you fail to comply with these terms and conditions. You shall, upon such termination of your rights, destroy all Product, Modified Animals, and components thereof in your control, and notify Sigma of such in writing.
5. You may not use the Product to support the filing of a patent application in any country in the world that contains claims directed to the Product or its uses.

For information on purchasing a license to this Product for purposes other than Licensed Research Use, contact your local Sigma Sales representative, who will refer you to the proper licensing representative, or in the USA call 800-325-3010.

JumpStart, ReadyMix, and DirectLoad are trademarks of Sigma-Aldrich Co. LLC
CompoZr is a registered trademark of Sigma-Aldrich Co. LLC
Nucleofection and Nucleofector are registered trademarks of Amaxa GmbH.
TransIT is a registered trademark of Mirus Bio Corporation.

CN,PHC 06/14-1