

## Product Information

### Human IGF-I R ELISA Kit

for serum, plasma, cell culture supernatant, and urine

Catalog Number **RAB0230**

Storage Temperature  $-20\text{ }^{\circ}\text{C}$

## TECHNICAL BULLETIN

### Product Description

The Human IGF-I sR ELISA (Enzyme-Linked Immunosorbent Assay) kit is an *in vitro* enzyme-linked immunosorbent assay for the quantitative measurement of human IGF-I sR in serum, plasma, cell culture supernatants, and urine. This assay employs an antibody specific for human IGF-I sR coated on a 96 well plate. Standards and samples are pipetted into the wells and IGF-I sR present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated anti-human IGF-I sR antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of IGF-I sR bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

### Components

1. Human IGF-1 R Antibody-coated ELISA Plate (Item A) - RAB0230A: 96 wells (12 strips  $\times$  8 wells) coated with anti-human IGF-I sR.
2. 20x Wash Buffer (Item B) - RABWASH4: 25 mL of 20x concentrated solution.
3. Lyophilized Human IGF-I R Protein Standard (Item C) - RAB0230C: 2 vials, recombinant human IGF-I sR.
4. ELISA 1x Assay/Sample Diluent Buffer A (Item D1) - RABELADA: 30 mL of animal serum with 0.09% sodium azide as preservative. For Standard/Sample (serum/plasma) diluent.
5. ELISA 5x Assay/Sample Diluent Buffer B (Item E1) - RABELADB: 15 mL of 5x concentrated buffer. For Standard/Sample (cell culture medium/urine) diluent.
6. Biotinylated Human IGF-I R Detection Antibody (Item F) - RAB0230D: 2 vials of biotinylated anti-human IGF-I R (each vial is enough to assay half microplate).
7. HRP-Streptavidin (Item G) - RABHRP5: 200  $\mu\text{L}$  of 300x concentrated HRP-conjugated streptavidin.

8. ELISA Colorimetric TMB Reagent (HRP Substrate, Item H) - RABTMB3: 12 mL of 3,3',5,5'-tetra-methylbenzidine (TMB) in buffered solution.
9. ELISA Stop Solution (Item I) - RABSTOP3: 8 mL of 0.2 M sulfuric acid.

### Reagents and Equipment Required but Not Provided.

1. Microplate reader capable of measuring absorbance at 450 nm
2. Precision pipettes to deliver 2  $\mu\text{L}$  to 1 mL volumes
3. Adjustable 1-25 mL pipettes for reagent preparation
4. 100 mL and 1 liter graduated cylinders
5. Absorbent paper
6. Distilled or deionized water
7. SigmaPlot software (or other software which can perform four-parameter logistic regression models)
8. Tubes to prepare standard or sample dilutions

### Precautions and Disclaimer

This product is for Research Use Only. Not for Use in Diagnostic Procedures. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

### Preparation Instructions

1. Bring all reagents and samples to room temperature (18–25  $^{\circ}\text{C}$ ) before use.
2. Sample dilution: If samples need to be diluted, Assay Diluent A (Item D) should be used for dilution of serum/plasma samples. 1x Assay Diluent B (Item E) should be used for dilution of culture supernatants and urine. The suggested dilution for normal serum/plasma is 2-fold.

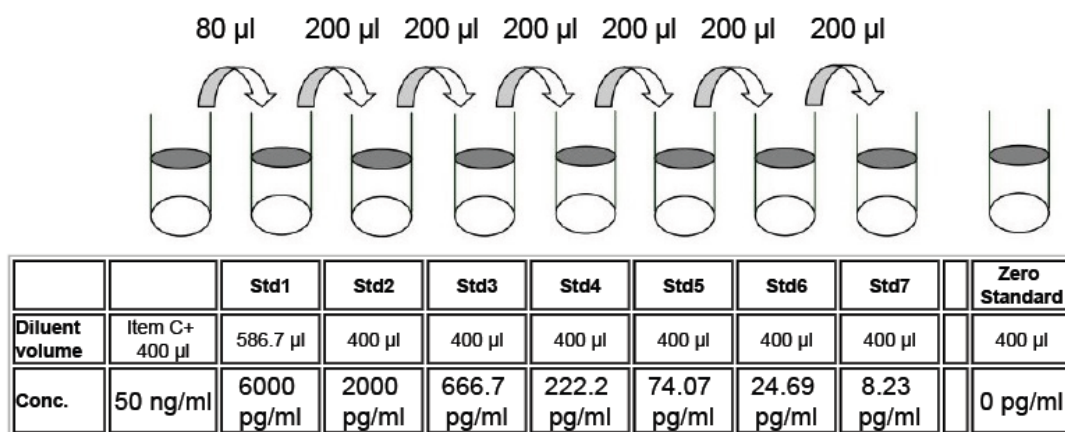
Note: Levels of IGF-I R may vary between different specimens. Optimal dilution factors for each sample must be determined by the investigator.

3. Assay Diluent B should be diluted 5-fold with deionized or distilled water.

4. Preparation of standard: **Briefly spin the vial of Item C** and then add 400  $\mu\text{L}$  of Assay Diluent A (for serum/plasma samples) or 1x Assay Diluent B (for cell culture medium and urine, Assay Diluent B should be diluted 5-fold with deionized or distilled water) into Item C vial to prepare a 50 ng/mL standard. **Dissolve the powder thoroughly by a gentle mix.**

Add 80  $\mu\text{L}$  of IGF-I sR standard from the vial of Item C into a tube with 586.7  $\mu\text{L}$  of Assay Diluent A or 1x Assay Diluent B to prepare a 6,000 pg/mL stock standard solution. Pipette 400  $\mu\text{L}$  of Assay Diluent A or 1x Assay Diluent B into each tube. Use the stock standard solution to produce a dilution series (see Figure 1). Mix each tube thoroughly before the next transfer. Assay Diluent A or 1x Assay Diluent B serves as the zero standard (0 pg/mL).

**Figure 1.**  
Dilution Series for Standards



5. If the Wash Concentrate (20x) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 mL of Wash Buffer Concentrate into deionized or distilled water to yield 400 mL of 1x Wash Buffer.
6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100  $\mu\text{L}$  of 1x Assay Diluent B into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4  $^{\circ}\text{C}$  for 5 days). The detection antibody concentrate should be diluted 80-fold with 1x Assay Diluent B and used in Procedure, step 5.

7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix gently before use. HRP-Streptavidin concentrate should be diluted 300-fold with 1x Assay Diluent B.

For example: Briefly spin the vial (Item G) and pipette up and down to mix gently. Add 40  $\mu\text{L}$  of HRP-Streptavidin concentrate into a tube with 12 mL of 1x Assay Diluent B to prepare a final 300-fold diluted HRP-Streptavidin solution (don't store the diluted solution for next day use). Mix well.

#### Storage/Stability

Store the kit at  $-20^{\circ}\text{C}$ . It remains active for up to 1 year. Avoid repeated freeze-thaw cycles.

The reconstituted standard should be stored at  $-20^{\circ}\text{C}$  or  $-70^{\circ}\text{C}$  ( $-70^{\circ}\text{C}$  is recommended). Opened microplate strips or reagents may be stored for up to 1 month at 2–8  $^{\circ}\text{C}$ . Return unused wells to the pouch containing desiccant pack and reseal along entire edge.

## Procedure

1. Bring all reagents and samples to room temperature (18–25 °C) before use. It is recommended that all standards and samples be run at least in duplicate.
2. Label removable 8 well strips as appropriate for the experiment.
3. Add 100  $\mu$ L of each standard (see Preparation step 4) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or overnight at 4 °C with gentle shaking.
4. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300  $\mu$ L) using a multichannel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100  $\mu$ L of 1x prepared biotinylated antibody (see Preparation, step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.
6. Discard the solution. Repeat the wash as in step 4.
7. Add 100  $\mu$ L of prepared Streptavidin solution (see Preparation, step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.
8. Discard the solution. Repeat the wash as in step 4.
9. Add 100  $\mu$ L of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.
10. Add 50  $\mu$ L of Stop Solution (Item I) to each well. Read at 450 nm immediately.

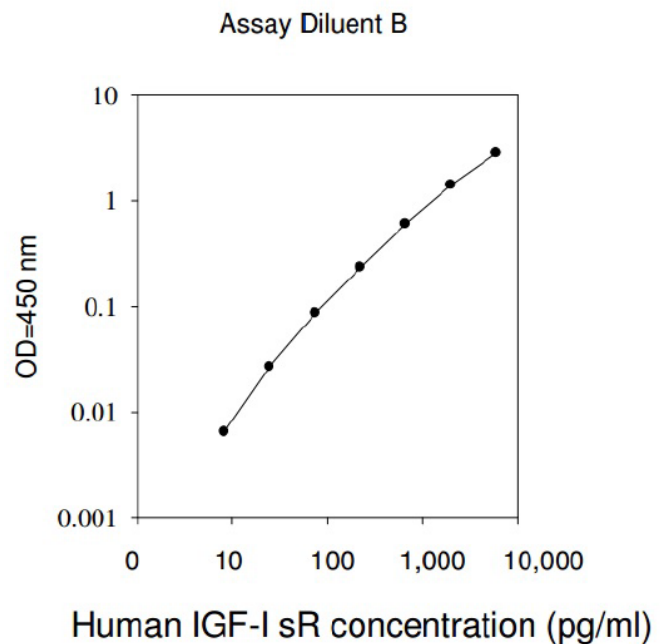
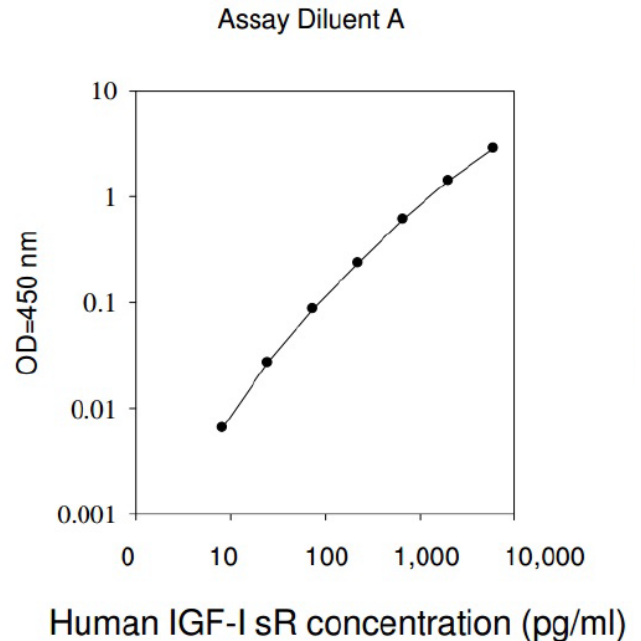
## Results

### Calculations

Calculate the mean absorbance for each set of duplicate standards, controls, and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

### Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



**Product Profile**

**Sensitivity:** The minimum detectable dose of Human IGF-I R was determined to be 6 pg/ml.

**Reproducibility:**

Intra-Assay: CV <10%

Inter-Assay: CV <12%

Minimum detectable dose is defined as the analyte concentration resulting in an absorbance that is 2 standard deviations higher than that of the blank (diluent buffer).

**Spiking & Recovery:** Recovery was determined by spiking various levels of human IGF-1 sR into human serum, plasma, and cell culture media. Mean recoveries are as follows:

Sample Type	Average % Recovery	Range (%)
Serum	92.48	78-103
Plasma	93.29	79-103
Cell culture media	94.12	80-104

**Linearity:**

Sample Type		Serum	Plasma	Cell Culture Media
1:2	Average % of Expected Range (%)	86 78-103	87 79-103	89 80-104
1:4	Average % of Expected Range (%)	91 81-104	89 80-104	91 82-105

**Specificity**

Cross Reactivity: This ELISA kit shows no cross-reactivity with any of the cytokines tested: Human Angiogenin, BDNF, BLC, ENA-78, FGF-4, IL-1alpha, IL-1beta, IL-2, IL-3, IL-4, IL-5, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12 p70, IL-12 p40, IL-13, IL-15, I-309, IP-10, G-CSF, GM-CSF, IFN-gamma, Leptin, MCP-1, MCP-2, MCP-3, MDC, MIP-1alpha, MIP-1beta, MIP-1delta, PARC, PDGF, RANTES, SCF, TARC, TGF-beta, TIMP-1, TIMP-2, TNF-alpha, TNF-beta, TPO, VEGF.

**Appendix**  
Troubleshooting Guide

<b>Problem</b>	<b>Cause</b>	<b>Solution</b>
Poor standard curve	Inaccurate pipetting	Check pipettes
	Improper standard dilution	Ensure a brief spin of Item C and dissolve the powder thoroughly with gentle mixing.
	Improper preparation of standard and/or biotinylated antibody	Briefly spin down vials before opening. Dissolve the powder thoroughly
Low signal	Too brief incubation times	Ensure sufficient incubation time; Procedure, step 3 may be done over night at 4°C with gentle shaking. Note: may increase overall signals including background.
	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation
Large CV	Inaccurate pipetting	Check pipettes
	Air bubbles in wells	Remove bubbles in wells
High background	Plate is insufficiently washed	Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed.
	Contaminated wash buffer	Make fresh wash buffer
Low sensitivity	Improper storage of the ELISA kit	Store the standard at <-20 °C after reconstitution, others at 4 °C. Keep substrate solution protected from light
	Stop solution	Add stop solution to each well before reading plate.

SA,ANK,KCP,KJ,MAM 09/18-1