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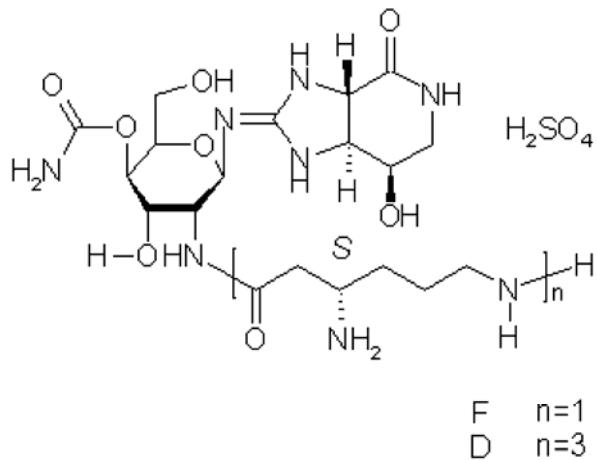
Nourseothricin sulfate BioChemika

Chemical name:

[3aS-[2(R^{*}), 3a α , 7a β]-2-[[4-O-(Aminocarbonyl)-2-deoxy-2-[(3,6-diamino-1-oxo-hexyl)-amino]- β -D-gulopyranosyl]amino]-3,3a,5,6,7,7a-hexahydro-7-hydroxy-4H-imidazo[4,5-c]pyridine-4-one dihydrogensulfate

Form:

Dihydrogensulfate of the weakly basic antibiotic Nourseothricin consisting of the components Streptothrin F and D

Structure:


Characteristics and use: In 1993, Nourseothricin has been introduced as a selection agent for molecular genetic research work. [1].

Nourseothricin is produced in cultures of a strain of *Streptomyces noursei*. [2]

Characteristic for Nourseothricin are the high stability of its crystalline salts (10 years at 4°C, 2 years at 20°C), its very good solubility in water, its relatively low oral toxicity (mice and other animals, no resorption in the gastrointestinal tract) and its wide range of antibiotic effects against gram-negative and gram-positive bacteria as well as against mycobacteria, mycoplasms, protozoa, certain DNA and RNA viruses and plants. The inhibition of the growth processes of yeasts and fungi is weaker. Nevertheless Nourseothricin sulphate is exceptionally suitable for the selection of recombinant yeast strains. [3].

The mechanism of action of Nourseothricin is comparable to that of other aminoglycoside antibiotics: Specific partial steps of protein synthesis are inhibited and miscoding is induced by the antibiotic. The development of resistance is based on monoacetylation of β -amino groups of the β -lysyl moiety of the Streptothrin molecules.

Nourseothricin sulphate is not used for therapeutic purposes. There is no cross resistance with drugs used in human and veterinary medicine. It is therefore very suitable as selection antibiotic in systems using resistance genes nat, sat, stat since resistance dominants, being able to cause nosocomial infections with medical relevant organisms, are not selected.

The plasmids pHN15 or pYL16, respectively, and Nourseothricin sulphate may be used as a selection system for microbial applications and for modern plant cultivation. The plasmids pHN15 and pYL16 contain the resistance genes nat 1 from *Streptomyces noursei*. The corresponding resistance genes sat1, sat2, sat3 originate from *Escherichia coli*, stat from *Streptomyces lavendulae*, encoding a Nourseothricin N-acetyltransferase.

The resistance genes are available as markers for heterologous expressions.

Nourseothricin may be used in fermentations of recombinant clones to maintain the permanent propagation of vectors carrying one of the resistance genes. [1, 2]

Solubility and storage of solutions: By dissolving 1000 mg Nourseothricine in 5 ml distilled water (material is not sterile filtered!) a stock solution of 200 mg/ml is yielded. The stock solution may be stored at +4°C for up to 4 weeks without detectable loss in activity. For longer storage of the solution up to 6 months, freezing at -20°C or lower is required.

Application notes [2]: Working solutions for use in molecular genetic applications are prepared by dilution of the stock solution with distilled water. Prior to use or even earlier, a sterile filtration is required. Recommended suitable Nourseothricin sulphate concentrations in nutrient media:

Escherichia coli	50 µg/ml
Saccharomyces cerevisiae	100 µg/ml
Ustilago maydis	75 µg/ml
Leishmania sp	>100 µg/ml
Cryptococcus neoformans	100 µg/ml
Arabidopsis thaliana	100 µg/ml

Safety

For R&D use only. Not for drug, household or other uses.

For safe handling kindly refer to our MSDS.

An addition of sodium hydroxide solution ($\text{pH} > 12$) inactivates Noureothricin sulphate within 3 hours by chemical degradation. [2]

References:

- H Krügel, G Fiedler, C Smith, S Baumberg, Sequence and transcriptional analysis of nourseothrycin acetyltransferase-encoding gene nat-1 from Streptomyces noursei, Gene 127, pp 127 – 131, 1993
- Supplier information
- A L Goldstein, J H McCusker, Three New Dominant Drug Resistance Cassettes for Gene Disruption in Saccharomyces cerevisiae, Yeast 15, pp 1541 – 1553, 1999
- H Bocker, F Bergter, Nourseothricin-Eigenschaften, Biosynthese, Herstellung, Arch exper Vet med, Leipzig, 40, pp 646 – 657, 1986
- S Horinouchi, K Furuya, M Nishiyama, H Suzuki, T Beppu, Nucleotide sequence of the streptothricin acetyltransferase gene from Streptomyces lavendulae and its expression in heterologous hosts, J Bacteriol 169, pp 1929 – 1937, 1987
- U Heim, E Tietze, W Weschke, H Tschäpe, U Wobus, Nucleotide sequence of a plasmid-born streptothricin-acetyltransferase gene (sat1), Nucleic Acids Res 17, p 7103, 1983
- H Krügel, G Fiedler, K Gase, I Haupt, Streptothricin resistance, in M E Bushell, U Gräfe (Eds): Bioactive metabolites from microorganisms, Progr Ind Microbiol, Amsterdam, 27, pp 357 – 367, 1989
- E Tietze, J Brevet, Nucleotide sequence of the streptothricin-acetyltransferase gene sat-2, Nucleic Acid Res 18, p 1290, 1990
- U Zähringer, W Voigt, G Seltmann, Nourseothricin (streptothricin) inactivated by plasmid pIE 636 encoded acetyl transferase of Escherichia coli. Location of the acetyl group. FEMS Microbiology Letters 110, pp 331 – 334, 1993
- K Smalla, R Prager, M Isemann, R Pukall, E Tietze, J.D. van Elsas, H Tschäpe, Distribution of streptothricin acetyltransferase encoding determinants among environmental bacteria. Mol. Ecol. 2, pp 27 – 33, 1993
- D. A. Chamberlain, R.I.S. Brettell, D.I. Last, B. Wirtzens, D. McElroy, R. Dolferus, E.S. Dennis, The use of the Emu promoter with antibiotic and herbicide resistance genes for the selection of transgenic wheat callus and rice plants. Australian J. Plant Physiol, pp 95 – 112, 1994
- S.E. Gild, G. Bakkeren, J.E. Davies, J. W. Kronstad, Three selectable markers for transformation of Ustilago maydis. Gene 142, pp 225 – 230, 1994
- P.B. Joshi, J.R. Davies, W. R. McMaster, The gene encoding streptothricin acetyltransferase (sat) as a selectable marker for Leishmania expression vectors. Gene, 156, pp 145 – 149, 1995
- M. Kojic, W. K. Holloman, Shuttle vectors for genetic manipulations in Ustilago maydis. Can. J. Microbiology 46, pp 333 – 338, 2000
- L. A. Zaslavkaia, J. C. Lippmeier, P. G. Kroth, A. R. Grossman, K. E. Apt, Transformation of the diatom Phaeodactylum tricornutum (Bacillariophyceae) with variety of selectable marker and reporter genes. J. Phycol. 36, pp 379 – 386, 2000
- J. Jelenska, E. Tietze, J. Tempé, J. Brevet, Streptothricin resistance as a novel selectable marker for transgenic plant cells. Plant Cell Reports 19, pp 298 – 303, 2000
- H. C. McDade, G.M. Cox, A new dominant selectable marker for use in Cryptococcus neoformans. Medical

- Mycology 39, pp 151 – 154, 2001
- H.Y. Steensma, J.J.M. Ter Linde, Plasmids with the Cre-recombinase and the dominat nat marker, suitable for use in prototrophic strains of *Saccharomyces cerevisiae* and *Kluyveromyces lactis*. Yeast 18, pp 469 – 472, 2001
- D. S. Fox, M. C. Cruz, R. A. L. Sia, H. Ke, G. M. Cox, M. E. Cardenas, J. Heitmann, Calcineurin regulatory subunit is essential for virulence and mediates interactions with FKBP12-FK506 in *Cryptococcus neoformans*. Molecular Biology 39 (4), pp 835 – 849, 2001
- G. Werner, B. Hildebrandt, W. Witte, Aminoglycoside-streptothricin resistance gene cluster assdE-sat4-aphA-3 Disseminated among multiresistand isolates of *Enterococcus faecium*. Antimicrob. Agents and Chemother 45 (11), pp 3267 – 3269, 2001
- K. Dornberger, H. Krügel, G. Peschel, HPLC determinations of the major factors of the nourseothricin complex, an antiviotic for non medial uses. J. Liq. Chrom. & Rel. Technol. 24(8), pp 1153 – 1159, 2001
- A. H. Y. Tong, M. Evangelista, A. B. Parsons, H. Xu, B. D. Bader, N. Pagé, M. Robinson, S. Raghbizadeh, Ch. W. V. Hogue, H. Bussey, B. Andrews, M. Tyers, Ch. Boone, Systematic genetic analysis with ordered arrays of yeast deletion mutants. Science, 294, pp 2364 – 2368, 2001
- R. Breitling, S. Klingner, N. Callewaert, R. Pietrucha, A. Geyer, G. Ehrlich, R. Hartung, A. Müller, R. Contreras, St. M. Beverley, K. Alexandrov, Non-pathogenic trypanosomatid protozoa as a latform for protein research and production. Prot. Express. Purific 25, pp 209 – 218, 2002
- N. Grammel, K. Pankevych, J. Demydchuk, K. Lambrecht, H.-P. Saluz, U. Keller, H. Krügel, A β -lysine adenylating enzyme and a β -lysine binding protein involved in poly β -lysine chain assembla in nourseothricin synthesis in *Streptomyces noursei*. Eur. J. Biochem. 269 pp 347 – 357, 2002
- R. C. Davidson, J. R. Blankenship, P.R. Kraus, M. de J. Berrios, Ch. M. Hull, C. D'Souza, P. Wang, J. Heitman A PCR-based strategy to generate inegrativ targetin alleles with large regions of homology, Microbiology 148, pp 2607 – 2615, 2002
- M. J. Hernandes-Lopez, A. Blascon, J.A. Prieto, Ura(-)host strains for genetic manipulation and heterologeous ecpressions of *Torulaspora delbrueckii*. Int. J. Food Microbiol. 88 (1-2), pp 79 – 86, 2003
- M. Teuber, F. Schwarz, V. Perreten, Molecular structure and evolution of the conjugativ multiresistance plasmid PRE25 of *Enterococcus faecalis* isolated from raw-fermented sausage, Int. J. Food Microbiol. 88 (2-3), pp 325 – 329, 2003
- T. Roemer, B. Jiang, J. Davison, T. Ketela, K. Veillette, A. Breton, F. Tandia, A. Linteau, S. Sillaots, C. Marta, N. Martell, S. Veronneau, S. Lemieux, S. Kauffman, J. Becker, R. Storms, Ch. Boone, H. Bussey, Large-scale essential gene identification in *Candida albicans* and applications to antifungal drug discovery. Molex. Microbiol 50 (1), pp 167 – 181, 2003
- J. Hansen, T. Felding, P.F. Johannesen, Further development of the cassette-bades pYC plasmid system by incorporation of the dominant hph, nat and AUR1-C gene markers and the lacZ reporter system, FFEMS Yeast research 4(3), pp 323 – 327, 2003
- A. Idnurm, J. L. Reedy, J. C. Nussbaum, *Cryptococcus neoformans* virulence gene discovery through insertional mutagenesis. Eukaryotic Cell 3(2), pp 420 – 429, 2004
- O. Reuss, A. Vikm R. Kolter, J. Morschhaeuser, The SAT1 flipper, an optimized tool for gene disruptions in *Candida albicans*, Gene 341, pp 119 – 127, 2004
- J. Shen, W. Gou, J. R. Koehler, CaNAT1, a heterologous dominant selectable marker for transformation of *Candida albicans* and other pathogenic *Candida* species. Infection and Immunity, 73 (2), pp 1239 – 1242, 2005